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Sources of short-lived bromocarbons in the Iberian upwelling system

S. Raimund¹, B. Quack², Y. Bozec¹, M. Vernet¹, V. Rossi³, V. Garçon³, Y. Morel⁴, and P. Morin¹

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Correspondence to: S. Raimund (raimund@sb-roscoff.fr)

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¹CNRS-UPMC Univ Paris 06, UMR 7144, Adaptation et Diversité en Milieu Marin, Equipe Chimie Marine, Station Biologique de Roscoff, 29682 Roscoff, France

²Leibniz Institute of Marine Sciences, IFM-GEOMAR, Kiel, Germany

³Laboratoire d'Etudes en Geophysique et Oceanographie Spatiales, CNRS, Toulouse, France ⁴Service Hydrographique et Océanographique de la Marine, (SHOM), Toulouse, France

Seawater concentrations of the four brominated trace gases dibromomethane (CH₂Br₂), bromodichloromethane (CHBrCl₂), dibromochloromethane (CHBr₂Cl) and bromoform (CHBr₃) were measured at different depths of the water column in the Iberian upwelling off Portugal during summer 2007. Bromocarbon concentrations showed elevated values in recently upwelled and aged upwelled waters (mean values of 30 pmol L⁻¹ for CHBr₃), while values in the open ocean were significantly lower (7.4 pmol L⁻¹ for CHBr₃). Correlations with biological variables and marker pigments indicated that phytoplankton could be identified as a weak bromocarbon source in the open ocean. In upwelled water masses along the coast, halocarbons were not correlated to Chl-a, indicating an external source, overlapping the possible internal production by phytoplankton. We showed that the tidal frequency had a significant influence on halocarbon concentrations in the upwelling and we linked those findings to a strong intertidal coastal source, as well as to a transport of those halocarbon enriched coastal waters by westward surface upwelling currents. Coastal sources and transport can be accounted for maximum values of up to 185.1 pmol L⁻¹ CHBr₃ in the upwelling.

Comparison with other productive marine areas revealed that the Iberian upwelling had stronger halocarbon sources than the phytoplankton dominated sources in the Mauritanian upwelling. However, the concentrations off the Iberian Peninsula were still much lower than those of coastal macroalgal influenced waters or those of polar regions dominated by cold water adapted diatoms.

1 Introduction

Oceans deliver short lived bromocarbons such as dibromomethane (CH₂Br₂), bromodichloromethane (CHBrCl₂), dibromochloromethane (CHBr₂Cl) and bromoform (CHBr₃) via exchange processes from the marine boundary layer to the atmosphere. Photooxidative breakdown of bromocarbons elevates atmospheric bromine

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concentrations and affects both tropospheric and stratospheric chemistry and has implications on the climate (Law and Sturges, 2006). In the troposphere, bromine influences the NO/NO₂ cycle, stimulates the catalytic ozone depletion cycles and influences the lifetimes of other trace gases (Read et al., 2008; von Glasow et al., 2004). Rapid deep convection can transport bromocarbons to the lower stratosphere and may contribute to 20-60% of stratospheric bromine and consequently contribute to ozone depletion there (Salawitch et al., 2005). Macroalgae and phytoplankton are the main sources for oceanic bromocarbons (Manley et al., 1992; Moore et al., 1996; Carpenter and Liss, 2000; Butler et al., 2007) and cause - depending on algae growth and species composition - high spatial and temporal variances (Carpenter et al., 2005) of the global halocarbon distribution. An important biological production pathway of polyhalogenated alkanes involves the catalysis by haloperoxidases (Leblanc et al., 2006) an enzyme found both in macroalgae and phytoplankton - but the biological purpose of these reactions still remains unclear. Besides natural sources, anthropogenic discharge contribute as a significant additional halocarbon source in some coastal areas (Quack and Wallace, 2003). Although a wide range of marine regions were investigated for aqueous bromocarbons, only a few studies focused on the highly productive and phytoplankton dominated upwelling regions. In laboratory experiments (Moore et al., 1996) and field campaigns (Hughes et al., 2009) haloperoxidases containing phytoplankton species were accounted for elevated halocarbon concentrations. Hence it was assumed that upwelling regions were important ecosystems for the global bromocarbon production. In this work we present seawater concentrations of CH₂Br₂, CHBrCl₂, CHBr₂Cl and CHBr₃ studied during late summer 2007 along the coast off Portugal and discuss different possible sources for these trace gases.

2.1 Sampling and study area

During summer 2007 (11 August to 14 September), we investigated halocarbon distributions in the Iberian peninsula upwelling system along the coast off Portugal onboard the French research vessel *Pourquoi pas?* during the MOUTON (*Modélisation océanique d'un théâtre d'opérations navales*) campaign. Samples were taken at 69 CTD stations along three different transects. Additional samples were taken during four daily cycles at four fixed stations (Fig. 1) with a 12-bottle CTD rosette (10-L-Niskin bottles). At each station, up to five samples were collected in the water column: surface, upper thermocline, maximum of chlorophyll, lower thermocline and bottom depth. Samples were taken to determine bromocarbon concentrations, pigment composition and nutrientconcentrations. Meteorological conditions and bio-physical data from this cruise are discussed in details by Rossi et al. (2010a).

The study area extended from 39.1 to 42.8° N and 8.9 to 11.1° W within the Iberian peninsula upwelling system, which is the northern extent of the wind driven Canary upwelling (eastern boundary upwelling systems). The Iberian peninsula upwelling system ranges from about 36° N to about 46° N and from the Iberian coast to roughly 24° W (Perez et al., 2001). During summer north/northwesterly tradewinds create a southward flow, which generates an offshore Ekman transport. The Ekman transport is responsible for the upwellingof cold and nutrient enriched waters from 100–300 m depth to the surface along the coastline (Smyth et al., 2001). Upwelling events are usually strongest in the north of the Iberian peninsula upwelling system (off Cap Finisterre) and are often related to westward flowing advection. These so called filaments are bands of cold and fresher waters which can reach as far as 100 km westward (Coelho et al., 2002).

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Environmental variables were recorded in the water column by a CTD rosette, as well simultaneously in the air by meteorological sensors of the vessel. Oxygen values were calibrated by independent sampling from discrete samples analyzed by Winkler titration. Pigments were analysed with the HPLC technique described by Wright et al. (1991), using a HPLC system (THERMO spectrasystem) equipped with a C18 (CLI) inverse column. One liter of sample was filtered at 200 MPa onto a 25 mm GF/F filter. Filters were stored at -196 °C in liquid nitrogen. Extraction of pigments was conducted with cold methanol (-20°C) for 12 h. Pigments identification and quantification was done by comparing retention times and peak areas and adsorption spectra obtained using certified standard solutions from DHI Group, Denmark.

Samples for nutrient analysis were taken in 125 ml polyethylene bottles, pre-treated with hydrochloric acid and deionised water, and finally rinsed with the sea water before sampling. Samples were stored at -20°C in darkness until the analysis was carried out. A semi-automated system (Bran & Luebbe, Autoanalyser II) was used to determine silicic acid (Si(OH)₄, nitrate (NO $_3$) and phosphate (PO $_4$).

Analysis of volatiles

Bromocarbons were analysed using a purge-and-trap technique and GC-ECD (Chrompack CP 9000) modified after Pruvost et al. (1999). The purge-and-trap-loop was altered and Valco valves were replaced by highly saltwater resistant Swagelok models. Sampling devices were modified according to Bulsiewicz et al. (1998). These new sampling devices (30 ml) were highly gastight and enable a straightforward connection to the Niskin bottle and to the purge-and-trap-loop respectively via Swagelok miniature quick connectors. The connector between the Niskin-bottle and sampling device comprises a filter element with 15 µm pore size, to remove larger particles. Samples were stored in the dark at 4°C and finally analyzed within four hours after sampling. Volatiles were extracted by purging with ultra-pure nitrogen for 20 min at a

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flow of 90 ml min⁻¹. Purging took place at ambient temperature in a purge chamber, which contained a glass frit (Pyrex 4). The gas flow was dried downstream using a condenser (held at 2°C) and a magnesium perchlorate trap. Volatiles were concentrated in a stainless steel capillary tube (150 cm) at -78°C and subsequently injected into a gas chromatographic column by thermodesorption (100°C, backflush). Separation of the compounds was performed using a capillary column (fused silica megabore DB-624, 75 m, 0.53 mm id, 3 mm film thickness, J & W Scientific, flow: 6 ml min⁻¹ ultra pure nitrogen) and a temperature program (10 min at 70°C, rising for 8 min to 150°C and stable for 7 min at 150°C). Quantification of volatiles was performed by external liquid standards (AccuStandard and Carlo Erba). Liquid standards were diluted in seawater and treated like a normal sample.

2.4 Statistics

Cluster analysis (k-means algorithm) of the data set revealed that SST forms three distinct clusters, which were used to separate the water masses. We defined the recently upwelled water mass with sampling stations showing a SST between 13.5 °C and 15.2 °C. These stations were close to the coast of the Iberian Peninsula. The aged upwelled water mass corresponds to stations with a SST between 15.5 °C and 17.7 °C. Those stations were located either close to the coast line or in the filament. The open ocean water mass featured a SST ranging between 18.1 °C and 20.8 °C. All these stations were located far off the Iberian coast. The three SST clusters were consequently used for all following data treatments (e.g. Pearson r correlations). We tested the effect of tide and time of the day on bromocarbon concentration using a one-way ANOVA with subsequent post-hoc tests (Tukey's honestly significant difference). For all statistical tests and techniques we used the software *Statistica* (Release 8.0).

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3.1 Upwelling during the campaign

The distribution of halocarbons in the Iberian peninsula upwelling system was one of the research topics of the MOUTON campaign in 2007. Water samples were taken in the upper 200 m of the water column along a north-to-south transect within the upwelling and along gradients from the coast to the open ocean. The sampling strategy was designed in due consideration of multidisciplinary interests, studying spatial and temporal variability of biological, physical and geochemical processes in the area. On board, satellite images were obtained for sea surface temperature (SST) and Chla concentration from the *Moderate Resolution Imaging Spectroradiometer* (MODIS aboard the NASA *Aqua satellite*). Those images were used for adjustment of the sampling strategy. In general, both Chl-a concentration- and SST-images indicated a proceeding upwelling and phytoplankton bloom with high daily variability along the coast off Portugal during all the campaign.

In order to describe the dynamics in the Iberian upwelling, we present the SST and Chl-*a* distributions, from satellite images taken on 19 August (Fig. 2). The SST-image shows clear upwelling in the studied area (Fig. 2a). Distinctive upwelling took place along the Iberian Peninsula from Cap Finisterre to 37° N with a larger intensity between 40° N and 42° N. A clear temperature gradient was visible from the open ocean (>18°C) towards the coast (<15°C). Pronounced cold water filaments occurred at 38° N, 40° N and 42° N, which showed significant temperature gradients with the surrounding water masses. These mesoscale structures (typical lifetime of a week) are created by advection processes and transport nutrient enriched cold water masses at the surface from the Iberian coast towards the open ocean. The Chl-*a* satellite images indicated elevated phytoplankton biomass along the Iberian Peninsula (Fig. 2b). Concentrations were highest near the coastline (above 2 mg m⁻³) and two orders of magnitude lower in the open ocean. Phytoplankton formed a meandering structure along the coast. During the campaign last week (4 to 9 September), upwelling conditions were still evident

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although wind velocity decreased (less than $10 \,\mathrm{m\,s}^{-1}$) and sea surface temperatures increased by several degrees along the coast.

The campaign started with a north-to-south track (Fig. 1), which followed the 100 m bottom depth isoline and was located within a zone of intense upwelling (14 to 17 August). This track was chosen to investigate the halocarbon concentrations dynamics within the maximum of phytoplankton density. A second transect (west-to-east track, Fig. 1) was performed along the 40° N latitude from the open ocean towards the upwelling (24 to 25 August). This track was chosen to investigate differences between coastal influenced upwelled waters and nutrient depleted open ocean waters. A distinct and newly developed offshore filament at 40.3° N with low SST values and high phytoplankton content was sampled from 6 to 9 September. Additionally, surface and depth samples were taken continuously at four stations (ST 1–4) during 30 h each: station 1 (40° N, 9.1° W; 18 to 19 August) and station 2 (41° N, 9° W; 20 to 21 August) were located in recently upwelled water masses. Station 4 (40.3° N, 9.2° W; 4 to 5 September) was chosen to investigate aged upwelled water masses. Station 3 (41° N, 10.5° W; 22 to 23 August) was a reference point in the open ocean.

3.2 Spatial distribution

Dibromomethane (CH₂Br₂), bromodichloromethane (CHBrCl₂), dibromochloromethane (CHBr₂Cl) and bromoform (CHBr₃) showed a very patchy distribution in the studied area (Fig. 3). Distribution patterns of the four measured brominated compounds were similar. Generally, concentrations were higher along the coast and lower in the open ocean as well as in the phytoplankton enriched cold water filaments. The highest coastal values were found between 41° N and 42° N and around 39.5° N. In these areas, surface concentrations were significantly elevated as compared to adjacent sampling stations and reached up to 185.1 pmol L⁻¹ for CHBr₃, 60.4 pmol L⁻¹ for CH₂Br₂, 17.5 pmol L⁻¹ for CHBr₂Cl and 6.8 pmol L⁻¹ for CHBrCl₂. In contrast, typical open ocean concentrations were about one order of magnitude lower than those maximum values.

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Cluster analysis of the data set revealed that SST forms three distinct clusters, which were used to classify different water masses (see methods). In order to evaluate processes in the biological active upper water layers, the clustered data set was additional divided into two groups (upper layers: surface to maximum of chlorophyll; lower lay-5 ers: below maximum of chlorophyll). Integrated values are presented in Table 1 for bromocarbons and complementary variables. Salinity showed slightly lower values of about 35.7 in the upwelling (both recent and aged upwelling) and elevated values in the open ocean (35.9). In addition, nutrients showed a clear decreasing concentration gradient, from the recently upwelled water masses to aged upwelled water masses to open ocean water masses. As a result of the amplified nutrient supply, both upwelled water masses feature higher pigment concentrations. For example, values for the diatom marker fucoxanthin were one order of magnitude higher in recently and aged upwelled waters as compared to values in open ocean waters. The spatial distribution of halocarbons in the biological active upper layers was characterized by a clear concentration gradient between both upwelled and open ocean water masses. Mean values were about 2 to 4.6 times lower in the open ocean as compared to upwelled waters. The high variability of halocarbon concentrations during the campaign becomes obvious in Fig. 4, showing mean values of the upper water layers for bromocarbons, SST, bottom depth (reflecting roughly the distance to the coast), Chl-a concentration and tidal amplitude. Bromocarbon concentrations were highest during the first week of the cruise, with significant elevated concentrations on the 15 to 17 August. The sampling site on the 15 August was located in the upwelling and influenced by the river plume discharge of the Minho and Douro, while the sampling site of the 17 August was in an active upwelling zone north of Lisbon. High bromocarbon concentrations were measured in recently and aged upwelled water masses, which are discernible by shallow bottom depth (reflecting near coast samples), elevated Chl-a concentrations and low SST. Samples from those water masses, however, also show high variability of bromocarbon concentrations, which did not correspond strictly to the distinct variances of SST- and Chl-a concentrations. A temporal variability is apparent in Fig. 4,

especially when evaluating the long duration sampling stations 1–4, where measurements were conducted during a daily cycle. The temporal variability will be discussed in more details with statistical tools in Sect. 4.3.

3.3 Vertical distribution of brominated halocarbons and environmental parameters

Concentrations of bromocarbons were generally elevated in the upper water layers (Table 1). Depth profiles of three representative stations – obtained during daily cycles – support this finding and provide a more detailed vertical resolution (Fig. 5). The variables Chl-a, N* and AOU are presented to evaluate whether phytoplankton could be a source of bromocarbons: Chl-a, as a general marker for the occurrence of phytoplankton, the apparent oxygen utilisation (AOU), which is influenced by biological oxygen production (photosynthesis) or consumption (respiration), and the quasi-conservative tracer N* (introduced by Gruber and Sarmiento, 1997), which is a linear combination of nitrate and phosphate. Since stratification has consequences on chemical and biological properties of the water column, density (σ) is the critical parameter for the evaluation of the vertical distribution of these parameters.

At station 2 (in the recently upwelled water mass), density revealed an intense mixing in the first 15 m without a clear pycnocline (Fig. 5b). Chl-a values were higher in the upper 25 m (1.4 mg m⁻³; Fig. 4a), followed by a moderate concentration gradient towards lowest Chl-a values below 43 m. AOU values were negatively correlated to Chl-a, with lowest values in the upper 25 m and high values in deep samples. N* displayed the same pattern as the AOU with a minimum in the upper 25 m (nitrate loss) followed by a clear gradient towards maximum values (nitrate production) below 43 m. The high Chl-a values, low AOU and low N* values observed in the upper 25 m were indicative of oxygen production and nitrate uptake likely due to dominant photoautotrophic processes. Below 43 m, the low Chl-a, high AOU, and high N* reflected considerable respiration by heterotrophic processes. In the biologically active upper water layers, bromocarbon concentrations were elevated and decreased constantly to

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low levels with depth. Among the four bromocarbons CH_2Br_2 and $CHBr_3$ showed a clear maximum at the depth of the Chl-a maximum. Daily variances of $CHBr_2Cl$ were high in comparison to the low concentration gradient between upper layer samples and lower layer samples.

At station 4 (in the aged upwelled water mass), the water column was stratified with a marked gradient from the surface to 33 m depth (Fig. 5f). The highest total Chl-*a* values for the Iberian upwelling were found in aged upwelled water masses (Fig. 4). At station 4, the highest Chl-*a* values were recorded at 20 m, together with low N* values (nitrate loss) and negative AOU values (photosynthetic oxygen production). Although we found the highest bromocarbon concentrations in the aged upwelled water masses, those values were restricted to only few sampling sites (Fig. 3). At station 4, bromocarbon values were lower that average upwelling concentrations (Table 1), but showed a typical gradient with low values in the deeper levels. For instance, the highest CHBr₃ concentrations at 20 m (12.4 pmol L⁻¹) were about three times higher than at 90 m but only the half of average upwelling concentrations (integrated in the upper layers).

At station 3 (in the open ocean), the water column was clearly stratified and showed a deep Chl-*a* maximum at 74 m (Fig. 5i–j). The pycnocline coincided with the highest phytoplankton density and activity (Chl-*a* maximum, N* minimum, low AOU; Fig. 5i–j). Bromocarbons concentrations were low compared to typical upwelling concentrations (Table 1). CHBr₃ concentrations were highest at the Chl-*a* maximum (5.7 pmol L⁻¹) at 74 m. The highest concentrations of CH₂Br₂ (5.3 pmol L⁻¹) and CHBrCl₂ (0.9 pmol L⁻¹) were measured between the Chl-*a* maximum and the end of the thermocline (from 74 to 110 m). CHBr₂Cl showed high daily variances and the highest values above the Chl-*a* maximum (1.7 pmol L⁻¹ at 51 m).

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Evaluating the source strength

In this study, bromocarbons concentrations reported for the first time in the Iberian upwelling were low compared to other productive marine areas (Table 2). To the best of our knowledge only three studies focused on upwelling regions. Class and Ballschmiter (1988) measured bromocarbons in air and water samples near the West African coast (6-30° N, 16° W). Quack et al. (2007) studied the oceanic distribution of CH₂Br₂ and CHBr₃ in the water column in the same area (17-20.5° N, 16-19° W). While CH₂Br₂ values were similar, reported CHBr₃ values were significantly lower in the latter study. A more recent work by Carpenter et al. (2009) presented sea surface concentrations for the same compounds, which were in close agreement with those from Quack et al. (2007). Both investigations agreed that bromocarbon concentrations were elevated in the upwelling, as compared to open ocean concentrations but lower than concentrations known from macroalgae influenced coastal regions. In agreement with results from the Mauritanian upwelling by Quack et al. (2007) and Carpenter et al. (2009), our results exhibit a marked gradient of bromocarbon concentrations from the upwelling towards the open ocean. Integrated values of CHBr₃ for the entire upwelling (30 pmol L⁻¹) were one order of magnitude lower than for other productive coastal areas (e.g. Jones et al., 2009; Bravo-Linares and Mudge, 2009) and hence do not support the fact that upwelling regions are pronounced "hot spots" for halocarbon formation, as previously assumed (Quack et al., 2004). However, sources for the Iberian upwelling seem to be higher than those for the Mauritanian upwelling since CH₂Br₂ and CHBr₃ values were significantly lower the off Mauritania coast.

The low bromocarbon concentrations measured in several upwelling areas questioned the putitative role of phytoplankton as an important halocarbon source and were in marked contrast with studies showing correlations between high concentrations of various halocarbons and the growth of microalgae (Krysell, 1991; Hughes et al., 2009;

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Klick and Abrahamsson, 1992; Moore and Tokarczyk, 1993). For example, CHBr₃ values reported by Carpenter et al. (2007) for phytoplankton rich Antarctic surface waters were about twice as much as in the Iberian upwelling. The discrepancy between expected and observed concentrations in upwelling areas investigated so far is likely 5 due to the fact that halocarbon production is highly species specific (e.g. Moore et al., 1996). Several authors showed that phytoplankton species from distant phyla are effective halocarbon sources and that within the same genus, species showed surprising low performance in halocarbon production (Sturges et al., 1992; Tokarczyk and Moore, 1994; Moore et al., 1996). Hill and Manley (2009) implicated halocarbon production with temperature adaptation of phytoplankton species: polar diatom species showed generally high bromoperoxidase activity, while most of the temperate and tropical species showed only little or no bromoperoxidase activity. Hence, phytoplankton composition in the investigated upwelling regions must have been unfavourable for a high bromocarbon production. It must be considered that differences in the published halocarbon concentrations in several areas may also be partly due to temporal variability of the source strength. Several studies showed that seasonality of halocarbon production can be explained by enhanced tissue decay during midsummer (Goodwin et al., 1997a; Klick, 1992), stimulated halocarbon formation under high light condition by macroalgae (e.g. Carpenter et al., 2005; Nightingale et al., 1995; Pedersen et al., 1996) or phytoplankton blooms (e.g. Hughes et al., 2009). Thus, the above comparison of different studies only roughly enables to identify tendencies of local source strengths.

4.2 Evidence for halocarbon production by phytoplankton

Although bromocarbon concentrations were comparatively low and thus the putitative role of phytoplanktonic production less important, we found evidences for a weak phytoplanktonic source in our studied area. Spatial (Figs. 2 and 3) and vertical distribution patterns (Fig. 5) of halocarbons and Chl-a showed similarities. Correlation analyses for brominated compounds with environmental variables suggest a phytoplanktonic source off shore the Iberian peninsula (Fig. 6). For the correlation analysis, the data set was

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clustered according to the water masses. Significant Pearson r correlations (p < 0.05) with environmental variables were plotted for each bromocarbon. Additionally to the biological markers N* (variable 1) and AOU (variable 2), we used light transmission (variable 3) as a rough approximation for phytoplankton density, since elevated phytoplankton densities induce a decrease in light transmission. Finally, correlations to marker pigments (variables 4–16) were used to identify phytoplankton groups for the production of halocarbons.

The most obvious observation is that correlations from bromocarbons with environmental variables were the strongest in the open ocean. The absence of clear correlations in recently upwelled and aged upwelled waters, together with elevated bromocarbon concentrations in these water masses, indicates a strong non-phytoplanktonic source, which possibly masked the halocarbon production by phytoplankton in these water masses. Consequently, phytoplankton had highest relevance as a bromocarbon source in the open ocean.

Negative correlations from CHBr₃ to the biological markers N* and AOU (variables 1 and 2) revealed that elevated bromocarbon concentrations were related to nitrate uptake and biological oxygen production, both typical for the occurrence of phytoplankton. Negative correlations to light transmission (variable 3) were also found for all bromocarbons in the open ocean. Assuming high phytoplankton densities caused a decreased light transmission, negative correlations to light transmission support the idea of a phytoplankton source. The diversity of correlated pigments emphasize the involvement of different phytoplankton groups in the formation of bromocarbons. Correlations were highest to fucoxanthin (marker for diatoms; variable 8), Chl-b (marker for green algae and for prochlorophytes; variable 5) as well as Chl-c3 and 19'-hexanoyloxyfucoxanthin (marker for prymnesiophytes; variables 7 and 11). Hence those plankton groups might have played a key role in the biological production.

Correlations of CHBr₂Cl and CHBrCl₂ to environmental variables were less pronounced than those of CHBr₃ and CH₂Br₂. Regardless of the purpose of halocarbon production (see Manley, 2002), the metabolic pathway within phytoplankton seems

to be preferable for CHBr₃ and CH₂Br₂. Nucleophilic substitution of CHBr₃ by chlorine ions could explain parts of the CHBr₂Cl and CHBrCl₂ production (Class and Ballschmiter, 1988). However it must be considered that halogen exchange reactions are extremely slow with an estimated CHBr₃ half-life time of 1.3 to 18.5 years (see references in Carpenter and Liss, 2000). Hence there will be a significant time delay between elevated CHBr₃ concentrations and chemical CHBr₂Cl and CHBrCl₂ production. Alternatively, the decay of organic matter, including the involvement of heterotrophic bacteria, would be an additional source for those bromocarbons. However, since AOU did not show positive correlations in any water mass the decay of organic matter must have been a relatively less important bromocarbon source.

4.3 Evidences for a near shore source

According to the results of the correlation analysis, phytoplankton dominated the production of brominated compounds in the open ocean. In the recently and aged upwelled waters however, phytoplankton production have been masked by strong non-phytoplanktonic or external sources. Such sources could be free floating macroalgae (Moore and Tokarczyk, 1993), decay of organic matter (Klick, 1992), or the transport of halocarbon enriched coastal waters to the open ocean (Carpenter and Liss, 2000; Chuck et al., 2005). Sources in coastal water are related to macroalgae (e.g. Nightingale et al., 1995) and, to a lower degree, to anthropogenic sources as discharge from power plants (Quack and Wallace, 2003).

A potential coastal source might explain the temporal dynamics of bromocarbon concentrations (Fig. 4). Hence we analysed the factors daytime and tide, which are known to have effects on coastal halocarbon concentrations (e.g. Carpenter et al., 1999). Analyses of variances (ANOVA) were computed for all samples taken from the biological active upper layers. For the ANOVA, the data set was arranged by the three water masses and additionally by time and tide, respectively. The factors time and tide were necessarily analysed separately, since the tidal cycles are synchronized to lunar month (27.3 day) intervals and hence daily tidal cycles shift forward in time. The tidal

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day is characterized by two high and two low water periods (semi-diurnal tide) along the Iberian coast. The temporal sequence of low and high tides are synchronized with the tidal amplitude: during the cruise, two spring tides with a maximum tidal amplitude occurred at 14 August and 31 August. At this period, the low tide passed around morning and afternoon. Then the low tide shifted constantly until the next neap tides (lowest tidal amplitude at 22 August and 5 September, respectively). At neap tide, the low tide passed around noon and midnight. Thus possible tidal and temporal effects on halocarbon concentration can complement or oppose one another.

The temporal division of the dataset followed the natural daily cycle and was clustered in four groups: night, morning, day and evening. Night and day were defined as two hours before sunrise and sunset, respectively. Morning and evening were defined as intermediates between day and night. During the campaign the day period lasted about 6.5 h, the night period about 9.5 h and morning and evening exactly 4 h in each case. The day time showed significant effects on most brominated halocarbon concentrations in recently upwelled water masses (Fig. 7). CH₂Br₂, CHBr₂Cl and CHBr₃ concentrations were significantly higher in the evening, as compared to the rest of the day (factor 1.5 to 1.8 at p-values below 0.055). In aged upwelled waters variances of bromocarbon concentrations were statistically not significant. In open ocean waters, diurnal variations of CH₂Br₂, CHBr₂Cl and CHBrCl₂ were also low and only CHBr₃ concentrations were significantly elevated during night (factor 2 at p = 0.05). Diurnal variations of halocarbon concentrations can be explained by elevated photon flux densities during midday, which boost the rate of photosynthesis and consequently enzymatic halocarbon production (Goodwin et al., 1997b). Such effects were shown for macroalgae (Ekdahl et al., 1998; Nightingale et al., 1995; Laturnus et al., 1998) and microalgae (Abrahamsson et al., 2004). The elevated midday concentrations in those studies are in contrast to our findings, where significant effects on CH₂Br₂, CHBr₂Cl and CHBr₃ concentrations were found in the evening. In order to investigate the factor tide and its potential impact on halocarbon transport, the data set was clustered in four groups: low tide, incoming tide, high tide and outgoing tide. The tidal clustering was

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achieved by coordinating sampling time with a tide table, provided by SHOM (Service Hydrographique et Océanographique de la Marine, Brest, France) for different locations along the Iberian coast. Hence the sampling time at sea reflected the tidal period at the coast. The influence of tide was evident for CH₂Br₂, CHBr₂Cl and CHBr₃ in recently upwelled waters, visible in aged upwelled waters, and not significant in the open ocean (Fig. 8). In recently upwelled waters, concentrations of CH₂Br₂, CHBr₂Cl and CHBr₃ were significantly elevated at outgoing tide (factor 1.6–1.9 at p < 0.01). In aged upwelled waters, tidal variations of these three compounds were higher with maxima at outgoing tide (2.2 to 2.3-fold elevated) but with a weaker significance level (0.05 > p > 0.1). Variances of CHBrCl₂ were statistically not significant in all water masses.

Our findings of general elevated bromocarbon concentrations at outgoing tide are in contrast to other studies, which previously investigated the effect of tide on halocarbon concentrations in air (Jones et al., 2009; Peters et al., 2005) and water (Nightingale et al., 1995; Carpenter et al., 2000) samples. In these studies, coastal air and water concentrations increased with lower water levels and showed minima at high water levels. As mentioned above our elevated concentrations found in the evening for several brominated compounds were in contrast to commonly found midday maximum. The discrepancies between our observations and previous studies reflect a possible coastal source associated with lateral transport of halocarbon enriched waters towards the upwelling.

The tidal and temporal effects on halocarbon concentrations in the upwelling can be explained by tidal mixing and upwelling currents. Tidal currents might elevate halocarbons concentrations at outgoing mid-tide. However those tidal residual currents are expected to be very low and variable at a dozen of km from the coast, the sampling location (Marta-Almeida and Dubert, 2006) and cannot explain the entire transport of water mass. Hence, the following mechanism is proposed. Intertidal macroalgae beds can be assumed as a source for elevated bromocarbons in the Iberian upwelling. The incoming tide is mixing and homogenizing near-shore waters, increasing their

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concentrations in freshly produced halocarbons. Then, the low tide associated with the westward surface upwelling currents (Rossi et al., 2010b) create an offshore transport of these bromocarbon enriched coastal waters. It induces elevated bromocarbon concentrations far from the coastline several hours after the low tide. The surface offshore velocities were ranging roughly from 0.1 to 0.2 m s⁻¹ for the first leg (strong upwelling favourable winds), giving a transport of 10 to 20 km (from the coast to the sampling sites) within 24 h. Hence high measured concentrations in the coastal upwelling might be related to an intertidal production during the previous tidal cycle, followed by an offshore transport.

Despite the general advection processes, bromocarbons were highly patchy distributed along the Iberian coast (Fig. 3). Causal for these dynamics can be different coastal sources (macroalgae, river discharge) with a high variability of either the source strength or the water transport towards the upwelling. The complex shelf topography of the Iberian upwelling System creates small scale instabilities, which may interfere with the transport caused by tidal mixing and upwelling currents. Along the northsouth transect, cyclonic eddies alternate with anticyclonic eddies (Rossi et al., 2010a). The small scale instabilities created highly variable patterns of cross-shore and along shore currents with effects on biological, physical and geochemical variables. Additionally it can be assumed that the source strength fluctuates along the Iberian coast, which may explain some particularly elevated bromocarbon concentrations between 41° N and 42° N and around 39.5° N. Lower salinities at those samples from the area indicate the presence of the Western Iberian Buoyant Plume (Peliz et al., 2002), which is characterized by important freshwater discharge from the rivers Minho and Douro. Hence anthropogenic discharge in the catchment area of the rivers could account for the unusual elevated bromocarbon concentrations. Alternatively macroalgae can be discussed as a possible source: areas of highest bromocarbon concentrations were coinciding with areas of the highest SST anomalies, computed by Relvas et al. (2007) for the period between 2001–2005. High SST anomalies determine elevated upwelling and consequently nutrient supply. We assume that this elevated nutrient supply might

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have positive implications on coastal macroalgae growth rates and consequently halocarbon production.

5 Conclusions

Our investigation of the Iberian peninsula upwelling revealed that CH₂Br₂, CHBrCl₂, CHBr₂Cl and CHBr₃ concentrations were elevated in the upwelling compared to the open ocean. A complex feature of along-shore and cross-shore currents and differences in source strength caused the strong patchiness of bromocarbon distribution in the studied area. Contrary to our assumption of a strong phytoplanktonic source within the upwelling, measured bromocarbon concentrations were low compared to macroalgae influenced coastal areas. Although the Iberian upwelling is thus not a pronounced "hot spot" for halocarbon formation, a weak phytoplanktonic source in the open ocean off Portugal was evident. For the upwelling along the coast however, the phytoplankton production was masked by strong coastal sources, including natural production by macroalgae and anthropogenic sources. The coastal sources showed a dependence on daytime and tide, a typical feature for halocarbon production in intertidal macroalgae beds.

We suggest that further studies should be conducted in the inner shelf between the shore line and the upwelling in order to localize important sources and assess the westwards transport of halocarbon enriched coastal waters. This type of study would complete our investigation of the middle and outer shelf of the Iberian upwelling and would provide a comprehensive understanding of halocarbon sources in this highly heterogeneous system.

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Table 1. The range (10th and 90th percentiles) and mean of bromocarbon concentrations and selected physical, chemical and biological variables in the Iberian Upwelling. Data grouped by sampling depth (upper layers: surface to maximum of chlorophyll; lower layers: below maximum of chlorophyll) and water mass with indicated temperature anomalies (difference between SST of a water mass and all samples).

	upper layers											
	recently upwelled				aged upwelled				open ocean			
	10th	90th	mean	N	10th	90th	mean	Ν	10th	90th	mean	Ν
CH ₂ Br ₂ [pmol L ⁻¹]	5.4	15.2	9.8	51	4.6	22.8	11.8	58	2.7	6.9	4.8	33
CHBrCl ₂ [pmol L ⁻¹]	1.3	3.8	3.1	50	0.9	3.9	2.3	57	0.5	1.0	8.0	33
CHBr ₂ Cl [pmol L ⁻¹]	0.6	5.1	3.2	51	0.0	8.2	3.3	58	0.5	2.2	1.3	33
CHBr ₃ [pmol L ⁻¹]	11.3	39.5	21.7	51	10.3	74.4	31.1	58	3.8	15.1	7.4	33
chl-a [mg m ⁻³]	0.57	2.57	1.40	52	0.37	3.48	1.75	73	0.13	0.90	0.50	33
$chl-b [mg m^{-3}]$	0.00	0.19	0.10	28	0.00	0.22	0.12	63	0.00	0.31	0.11	18
chl- c 2 [mg m $^{-3}$]	0.00	0.68	0.23	28	0.00	0.54	0.18	63	0.00	0.20	0.05	18
chl- c 3 [mg m $^{-3}$]	0.00	0.31	0.09	28	0.00	0.28	0.11	63	0.00	0.30	0.07	18
fuco [mg m ⁻³]	0.25	2.49	0.91	28	0.10	2.04	0.80	63	0.02	0.21	0.09	18
but [mg m ⁻³]	0.00	0.05	0.02	28	0.00	0.08	0.04	63	0.01	0.12	0.06	18
perid [mg m ⁻³]	0.03	0.15	0.09	28	0.02	0.22	0.10	63	0.00	0.11	0.03	18
hex [mg m ⁻³]	0.01	0.16	0.06	28	0.02	0.26	0.12	63	0.01	0.52	0.18	18
diadino [mg m ⁻³]	0.03	0.25	0.10	28	0.03	0.21	0.11	63	0.01	0.05	0.03	18
allo [mg m ⁻³]	0.00	0.06	0.02	28	0.00	0.05	0.02	63	0.00	0.02	0.01	18
lut [mg m ⁻³]	0.00	0.04	0.01	28	0.00	0.04	0.01	63	0.00	0.00	0.00	18
zea [mg m ⁻³]	0.00	0.06	0.03	28	0.00	0.09	0.04	63	0.05	0.11	0.08	18
bcar [mg m ⁻³]	0.00	0.13	0.05	28	0.00	0.11	0.04	63	0.00	0.06	0.02	18
Salinity [PSU]	35.64	35.72	35.68	52	35.61	35.76	35.69	73	35.80	36.04	35.92	33
Density $[\sigma]$	26.49	26.85	26.68	52	26.08	26.68	26.34	73	25.45	26.63	26.08	33
Oxygen [µmol L ⁻¹]	191.28	278.50	224.43	52	241.93	306.92	267.62	73	225.54	252.38	239.82	33
AOU [µmol L ⁻¹]	-21.87	63.64	32.91	52	-55.60	8.60	-14.25	73	-4.86	5.78	1.18	33
Nitrate [µmol L ⁻¹]	0.68	11.10	7.35	44	0.00	5.09	1.55	71	0.00	0.95	0.37	30
Silicate [µmol L ⁻¹]	0.70	5.83	3.13	45	0.54	1.97	1.05	72	0.91	1.94	1.27	31
Phosphate [µmol L ⁻¹]	0.11	0.97	0.62	44	0.05	0.38	0.18	70	0.02	0.17	0.08	33
N* [μmol L ⁻¹]	-2.11	1.87	0.19	43	0.33	2.24	1.36	69	1.46	2.80	2.07	30

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Table 1. Continued.

	lower layers												
	recently upwelled					aged upwelled				open ocean			
	10th	90th	mean	Ν	10th	90th	mean	N	10th	90th	mean	Ν	
CH ₂ Br ₂ [pmol L ⁻¹]	4.8	10.8	7.8	30	2.6	10.6	6.6	34	2.8	6.4	4.2	16	
CHBrCl ₂ [pmol L ⁻¹]	1.8	4.1	3.0	31	0.9	3.6	2.3	34	0.5	1.0	8.0	16	
CHBr ₂ CI [pmol L ⁻¹]	0.5	4.0	2.8	31	0.0	3.3	1.8	34	0.4	1.6	1.1	16	
CHBr ₃ [pmol L ⁻¹]	6.0	15.1	11.9	31	3.3	17.3	9.5	34	2.1	5.1	3.5	16	
chl-a [mg m ⁻³]	0.13	0.48	0.28	32	0.08	0.83	0.39	45	0.05	0.20	0.08	16	
chl-b [mg m ⁻³]	0.00	0.12	0.03	8	0.00	0.09	0.04	24				0	
chl- c 2 [mg m $^{-3}$]	0.03	0.17	0.07	8	0.00	0.10	0.03	24				0	
chl- c 3 [mg m $^{-3}$]	0.02	0.08	0.04	8	0.00	0.12	0.04	24				0	
fuco [mg m ⁻³]	0.13	0.72	0.33	8	0.05	0.55	0.23	24				C	
but [mg m ⁻³]	0.00	0.02	0.01	8	0.00	0.03	0.01	24				C	
perid [mg m ⁻³]	0.00	0.09	0.03	8	0.00	0.07	0.03	24				C	
hex [mg m ⁻³]	0.00	0.02	0.01	8	0.00	0.06	0.03	24				C	
diadino [mg m ⁻³]	0.01	0.07	0.03	8	0.00	0.04	0.03	24				C	
allo [mg m ⁻³]	0.00	0.06	0.01	8	0.00	0.01	0.00	24				C	
lut [mg m ⁻³]	0.00	0.02	0.00	8	0.00	0.03	0.01	24				0	
zea [mg m ⁻³]	0.00	0.04	0.01	8	0.00	0.03	0.01	24				C	
bcar [mg m ⁻³]	0.00	0.05	0.01	8	0.00	0.04	0.01	24				0	
Salinity [PSU]	35.67	35.82	35.76	32	35.70	35.82	35.77	45	35.72	35.80	35.76	1	
Density $[\sigma]$	26.92	27.05	26.99	32	26.80	27.05	26.94	45	26.90	27.05	27.01	1	
Oxygen [µmol L ⁻¹]	154.62	217.53	183.58	32	163.91	228.17	199.83	45	221.03	236.60	230.23	1	
AOU [μmol L ⁻¹]	52.26	107.21	80.14	32	1.31	98.91	54.90	45	24.65	43.11	33.73	1	
Nitrate [µmol L ⁻¹]	8.49	14.96	12.50	27	6.47	13.52	9.74	44	4.80	10.20	8.08	1	
Silicate [µmol L ⁻¹]	3.24	8.07	5.82	27	1.93	7.32	4.29	43	1.97	5.02	3.25	1	
Phosphate [µmol L ⁻¹]	0.60	1.03	0.86	27	0.45	0.92	0.68	43	0.31	0.71	0.53	1	
N* [µmol L ⁻¹]	-0.13	2.95	1.57	27	0.69	2.79	1.67	43	1.95	2.96	2.52	1	

chl-a (chlorophyll-a), chl-b (chlorophyll-b), chl-c2 (chlorophyll-c2), chl-c3 (chlorophyll-c3), fuco (fucoxanthin), but (19'butanoyl-fucoxanthin), perid (peridin), hex (19'-hexanoyl-fucoxanthin), diadino (diadinoxanthin), allo (alloxanthin), lut (lutein), zea (zeaxanthin), bcar (beta-carotene)

Table 2. Comparison of marine concentrations in different regions. Mean values in pmol L⁻¹.

	African coastal upwelling 25° N 16° W (03/1985) Class and Ballschmiter (1988)	Mauritanian upwelling 17–20.5* N 16–19* W (04–05/2005) Quack et al. (2007)	African coastal upwelling 16–35° N 14–24° W (05–06/2007) Carpenter et a	North east Atlantic 53–59° N 7–13° W (06–07/2006) II. (2009)	lberian Upwelling 39-43° N 9-11° W (08-09/2007) this work	English Channel 50° N 4° W (2006) Jones et al. (2009)	Irish Sea 53° N 4° W (2004–2005) Bravo-Linares and Mudge (2009)	Antarctic waters 70–72° S 9–11° W (12/2003) Carpenter et al. (2007)
CH ₂ Br ₂	5.77 ^a	4.9 ^a 5.8 ^b	3.4° 3 ^d 1.4°	1.1° 1.9 ^f 15.6 ^g	11.8 ^h 7.4 ⁱ	11.8 ^k 22 ^l	5.45 ^g	
$CHBrCl_2$	6.1ª				3.0 ^h 2.9 ⁱ		5.06 ^g	
CHBr ₂ Cl	9.6ª				3.7 ^h 2.6 ⁱ		17.3 ⁹	
CHBr ₃	23.74ª	10.7 ^a 9 ^b	11.5° 14.4 ^d 3.5°	3.4° 6.7 ^f 68.3 ^g	30.0 ^h 11.6 ⁱ	217.4 ^k 343.2 ^l	214.23 ^g	56.7 ⁹

^a depth 0–12 m; ^b depth 14–50 m; ^c upwelling; ^d Canary; ^e open ocean; ^f shelf and far coast; ^g coastal; ^h upwelling, upper layers; ⁱ upwelling, lower layers; ^k shore at high tide; ^l shore at low tide

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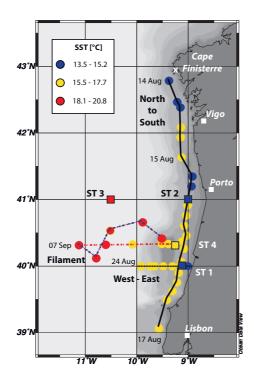


Fig. 1. Cruise track of the Mouton campaign 2007 in the Iberian Peninsula Upwelling System. Samples were taken at different CTD-stations (marked as dots) and along three different tracks (marked as lines). Solid line represents the north to south track within the upwelling. Dotted black line marks west to east track which followed 40° N from the open ocean toward the upwelling. Dashed red and blue lines show a track following a filament. Additional four stations were sampled during 30 h (ST 1-4; marked as squares). Colours represent station clustering by sea surface temperature (SST). Date specifications are given for selected stations.

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9°W

Longitude degree

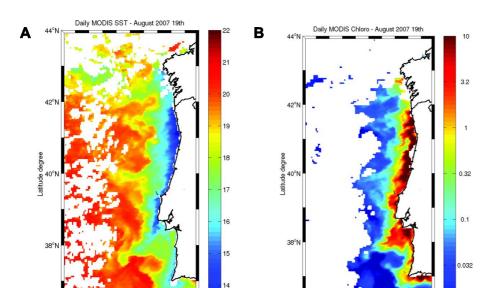


Fig. 2. Satellite images of sea surface temperature (SST) and Chl-*a* concentration on 19 August 2007. Source: Data from the Moderate Resolution Imaging Spectroradiometer (MODIS) aboard the Aqua satellite (EOS-PM1, NASA). **(A)** Blue reflects low SST (13°C), red high values (23°C). White is cloud cover. **(B)** Chl-*a* concentration [mg m⁻³].

13°W 12°W 11°W 10°W

36°N

12°W

10°W

11°W

Longitude degree

9°W

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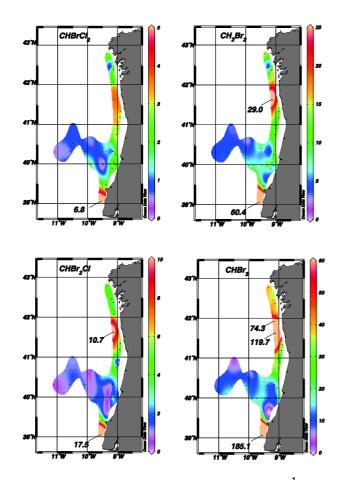


Fig. 3. Sea surface concentrations of bromocarbons in $pmol L^{-1}$. Bromodichloromethane (CHBrCl₂), dibromomethane (CH₂Br₂), dibromochloromethane (CHBr₂Cl) and bromoform (CHBr₃). Colour scales indicate different concentration ranges. Stations with values out of scale are labelled. Black dots are CTD stations.

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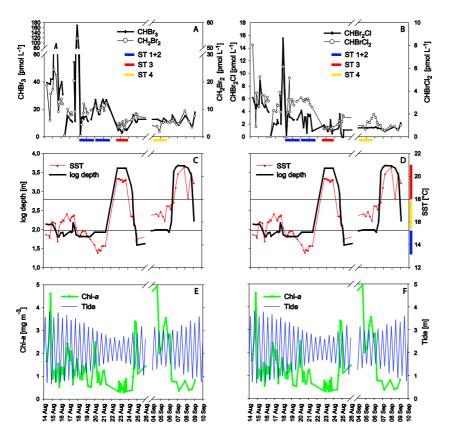


Fig. 4. Times series of bromocarbons and ancillary variables (mean values in the upper layers). (**A, B)** CH_2Br_2 , $CHBr_3$, $CHBr_2Cl$ and $CHCl_2Br$. Sampling at long observation stations 1–4 are marked with the colour code for the different water masses: Blue: recently upwelled waters. Yellow: aged upwelled waters. Red: Open Ocean waters. (**C, D)** bottom depth and SST. Lines are borders for the SST-cluster. Right axis shows the SST-range for the clusters. (**E, F)** Chl-a concentration and tidal amplitude.

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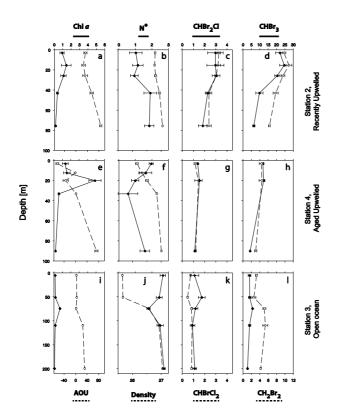


Fig. 5. Depth profiles of chlorophyll-a [mg m⁻³], AOU [μ mol L⁻¹], N* [μ mol L⁻¹], density [σ] and brominated halocarbons [pmol L⁻¹] at three long observation stations (ST 2-ST 4). Variables are expressed as means of all samples during a daily cycle at the different depths (surface, upper thermocline, maximum of chlorophyll, lower thermocline and bottom) and water mass: (a-d) Recently upwelled waters. (e-h) Aged upwelled waters. (i-l) Open ocean waters. Full lines use top-x-axis. Dashed lines refer to bottom-x-axis. Error bars show S.E.

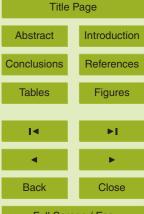


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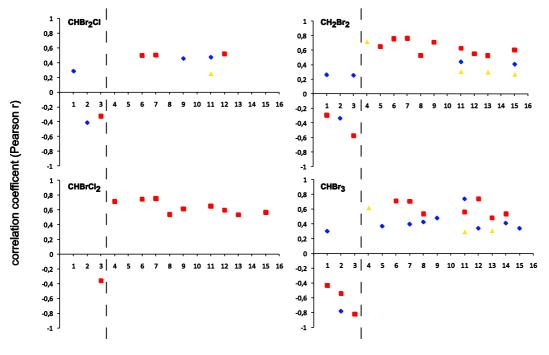


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environmental variables (legend see below)

Fig. 6. Results of cross-tabulation tables of Pearson r correlation coefficients for bromocarbons vs. environmental variables. Data are clustered into three water masses: Recently upwelled waters (blue), aged upwelled waters (yellow) and open ocean waters (red). Dashed line separates pigments from other variables. Variables 1-3: quasi biological markers (N*, AOU, light transmission). Variables 4–11: photosynthetic pigments (chl-a, chl-b, chl-c2, chl-c3, fuco, but, per, hex). Variables 12–16: photoprotective pigments (diadino, allo, lut, zea, bcar).

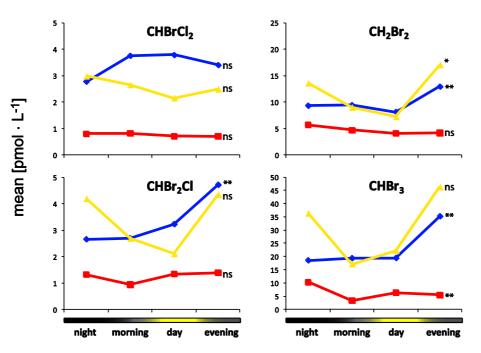


Fig. 7. Bromocarbon concentrations against time of the day in the upper water layers (surface to maximum of chlorophyll). Mean values at four different times of the day (night, day, and the intermediate morning and evening). Night and day were defined as two hours before sunrise and sunset, respectively. Blue lines: recently upwelled waters. Yellow lines: aged upwelled waters. Red line: open ocean waters. ns = not significant. * significant at 90%. ** significant at 95%.

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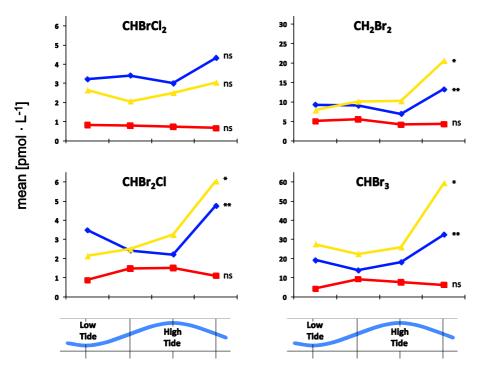


Fig. 8. Bromocarbon concentrations against tide in the upper water layers. Mean values of four tidal steps (low tide, incoming mid tide, high tide, outgoing mid tide). Blue lines: recently upwelled waters. Yellow lines: aged upwelled waters. Red line: open ocean waters. ns = not significant. * significant at 90%. ** significant at 95%.

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