

## Supplementary Information

# **Microbial community response to simulated petroleum seepage in Caspian Sea sediments**

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**Table S1.** Oligonucleotide probes used in this study

Probe name	Specificity	Formamide [%]	Sequence (5' - 3')	Reference
SCA1-212a	SCA-SRB1	20	CATCCCAAAACAGTAGCT	
SCA1-212b	SCA-SRB1	20	CATCCCCAACAGTAGCT	
h1_SCA1-197			TATWTATAGAGGCCA	
h2_SCA1-197			TATAWATAGAGGCCA	
h3_SCA1-182	Helper for SCA1-212ab		CCTTTGATCTRAAAA	Kleindienst <i>et al.</i> , 2014
h4_SCA1-182			CCTTTGATCTGAAWA	
h5_SCA1-229			GCTAATGGTACGCGRGCT	
h6_SCA1-182			CCTTTGATCTGGATA	
LCA2-63	LCA2	10	GC U A A G C U U U C U C G U U C	
h1_LCA2-83	Helper for LCA2-83		CUUUACUCACUCUAGCAA	Kleindienst <i>et al.</i> , 2014
Cyhx28-EdB_152	Clade Cyhx	20	ACGAAGCCTTCAGCATG	Jaekel <i>et al.</i> , 2015
Cyhx28-EdB_152_mod	Clade Cyhx	20	ACGAAGCCTTCGGCATG	This study
DSB985	<i>Desulfobacter,</i> <i>Desulfobacula,</i> <i>Desulfoviroa, Desulfotignum</i>	20	CACAGGATGTCAAACCCAG	Manz <i>et al.</i> , 1998
Arch915	Archaea	35	GTGCTCCCCGCCAATTCCCT	Stahl <i>et al.</i> , 1988
Delta495a			AGTTAGCCGGTGCTTCCT	
Delta495b	<i>Deltaproteobacteria</i>	30	AGTTAGCCGGCGCTTCCT	
Delta495c			AATTAGCCGGTGCTTCCT	
cDelta495a			AGTTAGCCGGTGCTTCTT	Loy <i>et al.</i> , 2002
cDelta495b	Helper for Delta495	30	AGTTAGCCGGCGCTTCKT	
cDelta495c			AATTAGCCGGTGCTTCTT	
Non338	Negative control	35	ACTCCTACGGGAGGCAGC	Wallner <i>et al.</i> , 1993
MS1414	<i>Methanoscinciales</i>	50	CTCACCCATACCTCACTCGGG	
hMS1395			GGTTTGACGGGCGGTGTG	Crocetti <i>et al.</i> , 2006
hMS1480	Helper for MS1414		CGACTTAACCCCCCTTGC	

**References**

- Crocetti, G., Murto, M., and Björnsson, L. (2006) An update and optimisation of oligonucleotide probes targeting methanogenic Archaea for use in fluorescence in situ hybridisation (FISH). *Journal of Microbiological Methods* **65**: 194-201.
- Jaekel, U., Zedelius, J., Wilkes, H., and Musat, F. (2015) Anaerobic degradation of cyclohexane by sulfate-reducing bacteria from hydrocarbon-contaminated marine sediments. *Frontiers in Microbiology* **6**: 116.
- Kleindienst, S., Herbst, F.A., Stagars, M., von Netzer, F., von Bergen, M., Seifert, J. *et al.* (2014) Diverse sulfate-reducing bacteria of the Desulfosarcina/Desulfococcus clade are the key alkane degraders at marine seeps. *ISME Journal* **8**: 2029-2044.
- Loy, A., Lehner, A., Lee, N., Adamczyk, J., Meier, H., Ernst, J. *et al.* (2002) Oligonucleotide Microarray for 16S rRNA Gene-Based Detection of All Recognized Lineages of Sulfate-Reducing Prokaryotes in the Environment. *Applied and Environmental Microbiology* **68**: 5064-5081.
- Manz, W., Eisenbrecher, M., Neu, T.R., and Szewzyk, U. (1998) Abundance and spatial organization of gram-negative sulfate-reducing bacteria in activated sludge investigated by in situ probing with specific 16S rRNA targeted oligonucleotides. *Fems Microbiology Ecology* **25**: 43-61.
- Stahl, D.A., Flesher, B., Mansfield, H.R., and Montgomery, L. (1988) Use of phylogenetically based hybridization probes for studies of ruminal microbial ecology. *Applied and Environmental Microbiology* **54**: 1079-1084.
- Wallner, G., Amann, R., and Beisker, W. (1993) Optimizing fluorescent in situ hybridization with rRNA-targeted oligonucleotide probes for flow cytometric identification of microorganisms. *Cytometry* **14**: 136-143.

**Table S2:** Pairwise comparison of community similarity within groups of samples based on presence-absence of A) bacterial and B) archaeal 16S rRNA OTU<sub>0.945</sub> of standardized data. Percentage of shared bacterial and archaeal OTU<sub>0.945</sub> between groups of samples is given.

A		SOFT 0-16 cm	untreated 4-8 cm	untreated 10-16 cm	SOFT 0-4 cm	SOFT 4-8 cm	SOFT 10-16 cm
Bacteria	untreated 0-16 cm	43					
	untreated 0-4 cm		36	31	56	38	42
	untreated 4-8 cm			33	39	51	37
	untreated 10-16 cm				33	36	61
	SOFT 0-4 cm					41	31
	SOFT 4-8 cm						35

B		SOFT 6-16 cm	untreated 10-16 cm	SOFT 6-10 cm	SOFT 10-16 cm
Archaea	untreated 6-16 cm	23			
	untreated 6-10 cm		21	59	18
	untreated 10-16 cm			18	60
	SOFT 6-10 cm				19

**Table S3:** Percentage of shared bacterial and archaeal 16S rRNA OTU<sub>0.945</sub>.

		No. of samples	Max shared OTU <sub>0.945</sub> (%)*)	Mean shared OTU <sub>0.945</sub> (%)*)	Min shared OTU <sub>0.945</sub> (%)*)
Bacteria	untreated	6	97	56	32
	SOFT	5	96	54	31
	0 – 4 cm	3	75	71	70
	4 – 8 cm	4	75	63	54
	10 – 16 cm	4	75	67	57
Archaea	untreated	5	83	58	39
	SOFT	5	80	60	38
	6 – 10 cm	5	79	61	41
	10 – 16 cm	5	74	65	45

\* Pairwise comparison of community similarity within groups of samples based on presence-absence OTU<sub>0.945</sub> of standardized data (resampling without replacements of 2730 sequences for bacterial 16S rRNA and 3908 sequences for archaeal 16S rRNA). Values refer to maximum, mean and minimum shared OTU<sub>0.945</sub> between any given pair of samples from the respective group.

**Table S4.** Relative abundance of 16S rRNA gene sequences retrieved from Caspian Sea untreated and SOFT sediments. Only sequences classified as *Desulfobacterales* by the SILVA NGS pipeline (release 119.1) were considered for further detailed phylogenetic analysis using arb.

Depth [cm]	Total <i>Desulfo- bacterales</i>	SCA1	C2-C4 alkane degr.*	SEEP1a	SEEP1b	SEEP1d	<i>Desulfo- sarcina</i>	<i>Desulfo- coccus</i> Hxd3	LCA1	LCA2	SB-29 relatives	s2551	Cyhx <sup>§</sup>	<i>Desulfo- bacula</i>	<i>Desulfati- glans</i> group	Sva0081	
Untreated	0-1	14,0	0,6	0,1	0,1	0,3	1,0	0,9	0,2	0,1	0,2	0,4	0,7	0,2	2,3	0,9	2,5
	2-4	8,3	0,3	<0,1	0,2	0,2	0,8	0,4	0,2	0,1	0,1	0,3	0,4	0,1	0,9	0,9	2,3
	4-6	12,8	0,6	0,1	0,2	0,2	1,6	0,6	0,3	0,1	0,2	0,6	0,5	0,2	0,9	1,1	4,5
	6-8	15,5	0,7	<0,1	0,3	0,6	1,1	0,4	0,2	0,1	<0,1	0,2	0,4	0,1	1,7	1,8	5,4
	10-12	13,1	0,6	<0,1	0,4	0,3	1,6	0,3	0,1	0,1	0,1	0,1	0,5	0,1	1,1	1,6	4,6
	14-16	19,4	0,6	0,1	0,4	0,9	3,4	0,5	0,3	0,2	0,3	0,3	0,5	0,3	2,0	1,8	5,7
SOFT	0-1	9,2	0,6	0,1	0,2	<0,1	0,1	0,5	0,1	<0,1	<0,1	0,3	0,5	0,5	2,4	0,9	1,8
	2-4	14,0	0,5	1,1	0,2	0,2	1,2	1,3	0,2	<0,1	0,8	0,4	0,5	0,6	2,7	1,3	3,1
	4-6	20,6	0,7	0,4	0,1	0,2	0,7	1,5	0,2	0,1	1,5	0,5	0,9	0,2	8,1	1,1	3,4
	6-8	22,3	0,8	0,7	0,3	0,4	2,7	0,6	0,2	0,1	1,9	0,2	0,7	0,4	6,4	2,1	4,2
	10-12	20,0	0,6	0,1	0,4	0,6	3,3	1,0	0,3	0,1	0,2	0,2	1,1	0,1	2,9	1,7	4,9
	14-16	15,9	0,8	0,1	0,2	0,5	3,4	0,5	0,3	<0,1	0,1	0,4	0,8	0,1	1,2	2,1	3,6

**Table S5.** Frequencies of archaeal 16S rRNA gene sequences retrieved from initial and SOFT core sediments (6 – 16 cm depth) that are affiliated with known taxonomic clades involved in the methane cycle. Total number of quality-trimmed archaeal 16S rRNA tag sequences: 25968 for untreated sediments and 128093 for SOFT sediments.

Taxonomy according to ARB SILVA (release 119).

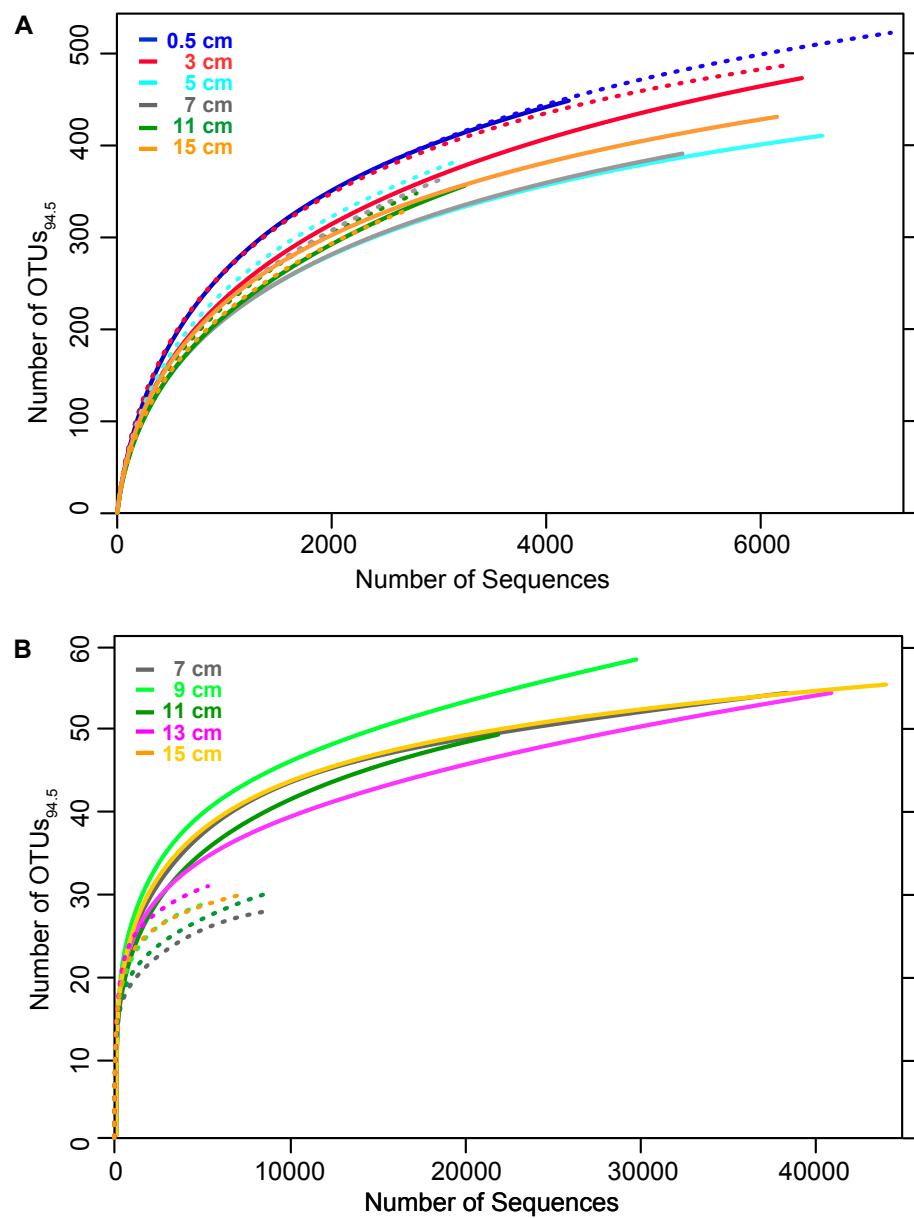
	Untreated sediment	SOFT sediment
	[% archaeal sequences]	
ANME-2a-2b		0.015
ANME-2c		0.001
GoM-Arch87		<0.001
Methermicoccaceae		0.009
Methanosaetaceae	0.004	0.012
Methanocellaceae		0.002
Methanomicrobiaceae		0.702
Methanosarcinaceae		
<i>Methanolobus</i>	0.065	0.126
<i>Methanococcoides</i>	0.008	0.040
<i>Methanosarcina</i>	0.004	11.125
<i>Methanosaeta</i>	0.004	0.016
Others (ANME-3, <i>Methanohalophilus</i>		0.020
<b>SUM [% of all Archaea]</b>	<b>0.085</b>	<b>12.069</b>



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**Figure S1**

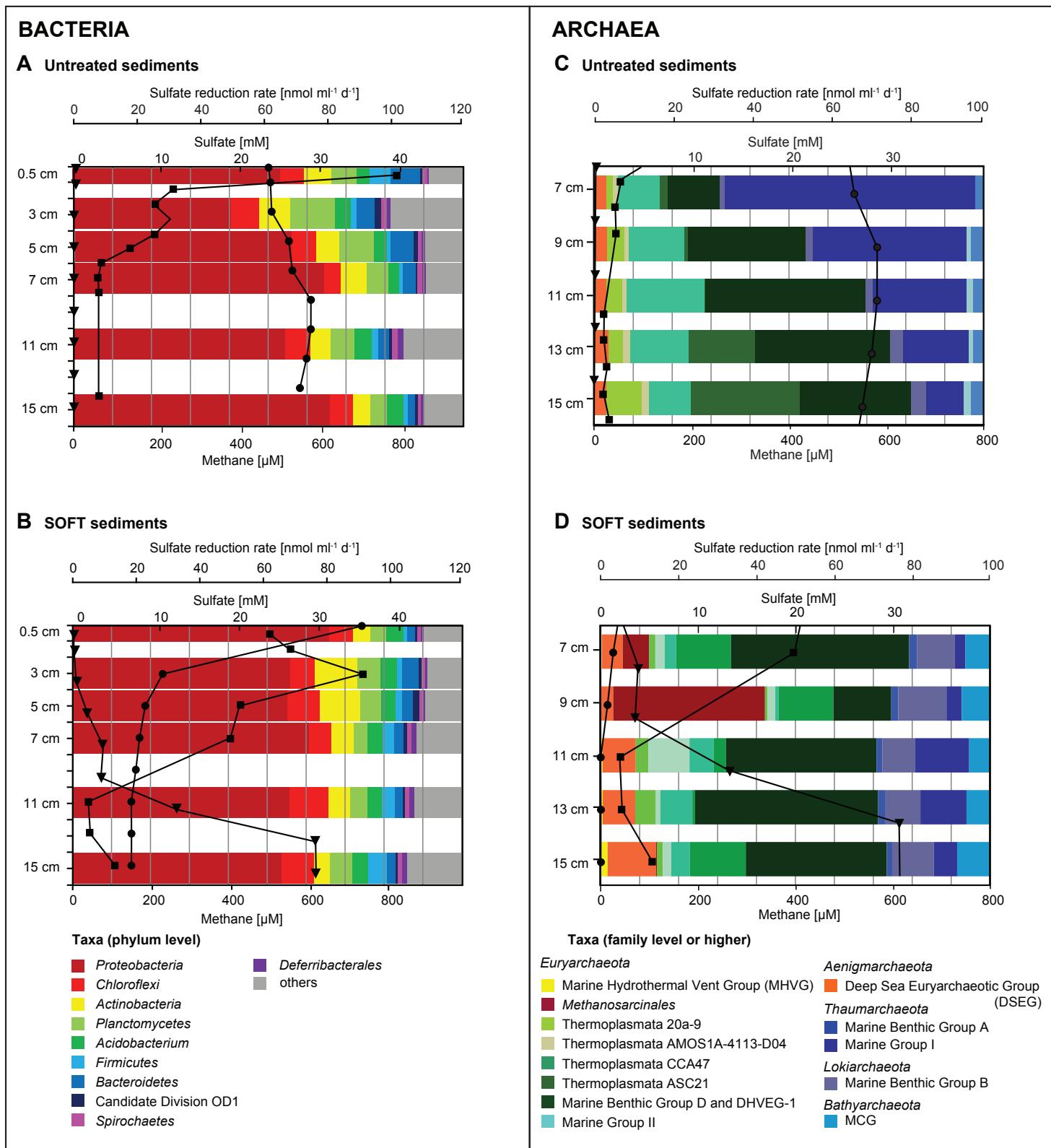
Map of the Caspian Sea showing major oil fields (white dots) and the sampling site off-shore Baku (Azerbaijan; red asterix).



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**Figure S2. Rarefaction curves**

Rarefaction curves for A) bacterial 16S rRNA sequences and B) archaeal 16S rRNA gene sequences clustered at 94.5% identity retrieved from SOFT (solid lines) and untreated (dashed lines) Caspian Sea sediment samples.



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**Figure S3.** Microbial community composition of Caspian Sea sediments in untreated (panels A, C) and SOFT (panels B, D) sediments. Relative abundance of (A, B) bacterial taxa (based on 454-pyrosequencing of 16S rRNA genes) and (C, D) archaeal taxa (based on IonTorrent-sequencing of 16S rRNA gene) is shown. Depth profiles for methane (triangles), sulfate (dots) and sulfate reduction rates (rectangles) were taken from Mishra *et al.* (this issue).