



Originally published as:

Qu, Y., Wu, N., Guse, B., Makarevičiūtė, K., Sun, X., Fohrer, N. (2019): Riverine phytoplankton functional groups response to multiple stressors variously depending on hydrological periods. - *Ecological Indicators*, 101, pp. 41—49.

DOI: <http://doi.org/10.1016/j.ecolind.2018.12.049>

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Riverine phytoplankton functional groups response to multiple stressors variously depending on hydrological periods

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18 **ABSTRACT**

19 Rivers and related freshwater ecosystems are facing increasing natural disturbance and
20 anthropogenic stressors. Understanding the key ecological processes that govern the riverine biota in
21 aquatic ecosystems under multiple pressures has crucial importance. However, there is still
22 insufficient knowledge in quantifying of stressors interactions. Moreover, the understanding of the
23 responses of riverine phytoplankton to multiple stressors is still scarce from catchment aspect. As an
24 interdisciplinary study, the catchment hydrological processes were linked to ecological responses in
25 this study, and we chose phytoplankton functional groups (PFGs) instead of taxonomic
26 classifications of algae to examine their responses to land-use pattern (L), hydrological regime (H),
27 and physicochemical condition (P) across two contrasting hydrological periods (dry, wet). The
28 traits-based phytoplankton functional groups are highly suggested as robust bio-indicators for better
29 understanding the current ecological status. The hydrological regime was described by a matrix
30 indices of hydrological alteration based on the outputs of a well-established ecohydrological model
31 (SWAT). The results from variation partitioning analysis showed that P and H dominate during the
32 dry period and P in high flows. Structural equation models (SEM) showed that the skewness of 7
33 days discharge emerged as a key driver of H, and had always an indirect effect on functional group
34 **TB** (benthic diatoms) during both hydrological periods. The functional group **M** (mainly composed
35 by *Microcystis*) has directly related to phosphorous in both periods, while indirectly to L of urban
36 area in high flow period, and water bodies in low flow period. This study emphasized that climate
37 change and anthropogenic activities such as altering flow regime and land-use pattern affect directly
38 or indirectly riverine phytoplankton via physicochemical conditions. In addition, our findings

39 highlighted that biomonitoring activities require detailed investigation in different hydrological
40 periods. SEM is recommended for improved understanding of phytoplankton responses to the
41 changing environment, and for future studies to fulfill the increasing demand for sustainable
42 watershed management regarding aquatic biota.

43 *Keywords:*

44 Phytoplankton functional groups,

45 Land-use pattern,

46 Hydrological regime,

47 Physicochemical condition,

48 Structural equation model

49

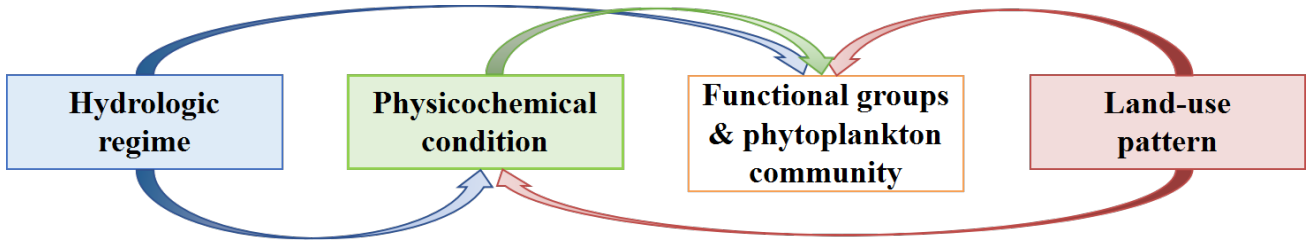
50 **1. Introduction**

51 Riverine phytoplankton is one of the vital primary producers in the river ecosystem, and acts as
52 robust bio-indicator responding to multiple stressors (Hilton et al., 2006). It is widely used for
53 bio-assessment of freshwater ecosystems in recent years (EC, 2000). Benthic diatoms are one of
54 significant elements of riverine phytoplankton recruitment, especially in the fine substrate rivers
55 (Bolgovics et al., 2017). Short water residence time lead to benthic diatoms dominance (Reynolds,
56 1994; Wang et al., 2018), while dynamic flow velocity results in shifts of the share of benthic
57 diatoms in the phytoplankton community (Wu et al., 2007). Moreover, flow alteration influence
58 riverine phytoplankton assemblages by altering nutrient delivery, light availability, dewatering of
59 habitats or severe vertical mixing (Reynolds, 2006). Despite diatom dominance, several studies
60 demonstrate that running water can also suffer the negative consequences of cyanobacterial blooms,
61 resulting from anthropogenic eutrophication and changing environment (Bowling et al., 2016;
62 Stanković et al., 2012). Intensive agriculture and urbanization strongly affect freshwater biodiversity
63 through flow modification and nutrient over-enrichment by fertilizer, pesticides, and sewage fluxes
64 (Bussi et al., 2016; Harris and Smith, 2016; Katsiapi et al., 2012). Changing climate conditions,
65 specifically global warming and altered rainfall patterns, play additional interactive roles in
66 modulating cyanobacterial blooms frequency, intensity and geographic distribution in a long-term
67 aspect (Paerl, 2017; Paerl et al., 2011). Therefore, multiple stressors including natural disturbances
68 (drought and floods) and anthropogenic stressors (human-induced water pollution, eutrophication)
69 are affecting riverine phytoplankton directly or indirectly. Understanding the key ecological
70 processes that govern riverine phytoplankton community in aquatic ecosystems under multiple

71 pressures has crucial importance. It is a fundamental prerequisite for robust bio-assessment, as well
72 as sustainable watershed management. However, investigations and analysis of stressors interactions
73 were much insufficient. Studies about the response of riverine phytoplankton to multiple stressors
74 from catchment scale are still scarce.

75 Interactions of multiple stressors determine the occurrence and survival of algae. In turn,
76 phytoplankton response to stressors by associated functions of tolerance, preference, and sensitivities
77 of distinct traits (Kruk et al., 2017; Litchman and Klausmeier, 2008). In this study, we chose
78 phytoplankton functional groups (PFGs) instead of taxonomic classifications of phytoplankton
79 dynamics to investigate their response to multiple stressors in a better way: land-use pattern (L),
80 hydrological regime (H) and physicochemical condition (P) (see Fig. 1, a schematic diagram
81 developing a conceptual framework). PFGs concept suggested aggregating species with similar
82 features into a few functional groups. Each group was described according to the physiological,
83 morphological and ecological traits, as well as common environmental sensitivities and tolerances
84 (Borics et al., 2007; Reynolds et al., 2002). For elaboration of the phytoplankton-based quality
85 assessment, Borics et al. (2007) replenished and evaluated PFGs in rivers, considering four parts:
86 trophic state, turbulence character, time sufficient for development of the given assemblage and risk.
87 Code **TB** (composed by benthic diatoms) and code **M** (mainly composed by limnophilic *Microcystis*
88 *spp.*) represent two highly different functional groups of riverine phytoplankton. **TB** is assigned to
89 mesotrophic, highly lotic preference benthic species with low risk of harm, while **M** assigned to
90 hypertrophic nutrient status, lentic preference, climax assemblages with potential toxicity. Mischke
91 et al. (2011) also selected the proportions of Cyanobacteria and Pennales (as typical benthic diatom)
92 to assess the trophic status of German Rivers. They concluded an unhealthy status with a high

93 proportion of Cyanobacteria, while they detected a healthy status with a high percentage of Pennales.
94 We were working with a lentic-lotic continuum watershed in a rural area under natural hydrology
95 disturbance and human-induced stressors. Based on the previous investigation of this catchment, we
96 found nutrients, especially the nitrate loads, were highly related with agriculture activities in the
97 Treene basin (Haas et al., 2016). **TB** were dominant in most of the study area, while their share
98 varied in time and space(Qu et al., 2018b). The temporary intensive occurrence of *Microcystis* in dry
99 season received high concerns from the local stakeholders (Qu et al., 2018a). Therefore, we are
100 especially interested in these two functional groups **TB** and **M**, in addition to the phytoplankton
101 community characteristics. We hypothesized that: 1) High flow condition promote to **TB** domination
102 over the others; 2) High share of agriculture land-use rise ascendancy of **M** in the community. Our
103 study aims to disentangle the causal-effects relationship of multiple stressors and riverine
104 phytoplankton community (indicated by PFGs) from wet and dry seasons. The answers would give
105 novel insights into the complex pathways of joint impact of natural and anthropogenic descriptors on
106 riverine phytoplankton community in lowland rivers across contrasting hydrological periods.



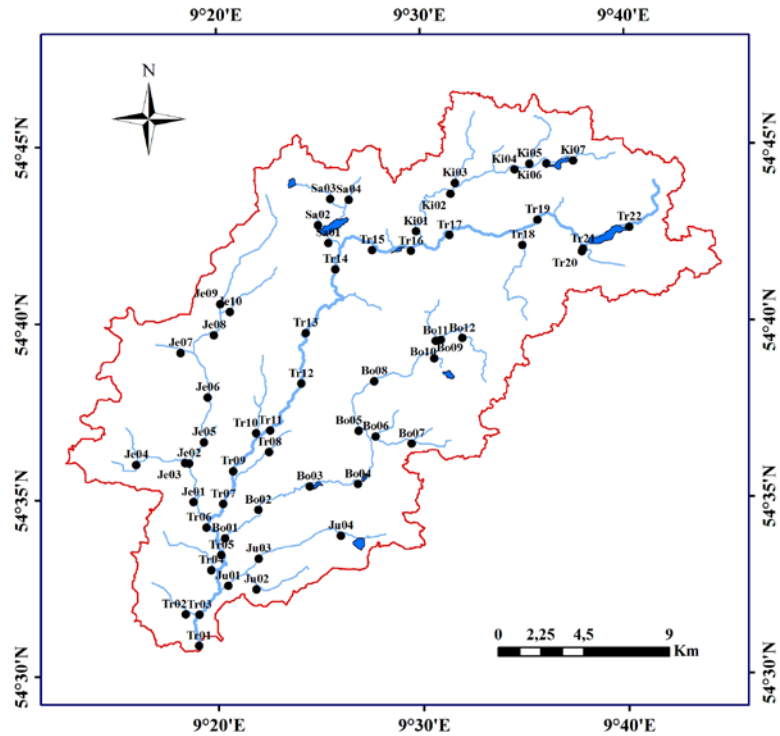
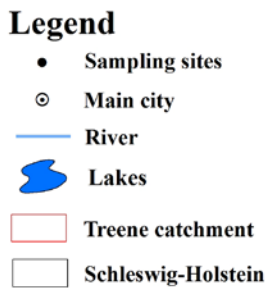
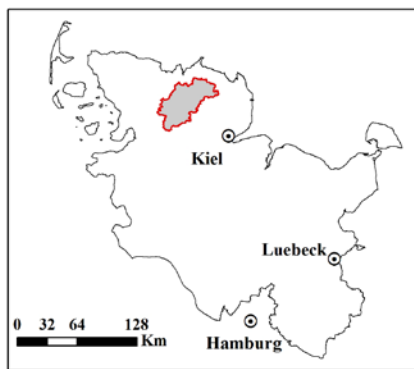
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108 **Fig. 1** Schematic diagram showing the major factors (e.g., hydrological regime, physicochemical condition, and land-use
109 pattern), which govern the pattern of phytoplankton functional groups and community structures. Those controls include
110 state factors, interactive controls, direct controls and indirect controls, which eventually causes the difference of
111 phytoplankton communities.

112

113 **2. Material and methods**

114 *2.1. Study area and sampling sites*

115 This study was carried out in Treene River, a lowland watershed located in the northern
116 Germany. As a lowland catchment, the maximum elevation of Treene basin is 76 m, with watershed
117 area of 517 km². The Treene catchment is in a temperate climate zone, which is influenced by marine
118 climate. Meanwhile it has a typical seasonal pattern of discharge, with the highest water flow in
119 winter and the lowest in summer and autumn (Guse et al., 2015a). Field surveys were carried out in
120 two hydrological periods (a wet period in December 2014 and a dry period in September 2015) on 59
121 sampling sites which covered mainstream and tributaries of the catchment. The abbreviation of the
122 site names are according to each sub-basin where they are located in: Bo for Bollingstedter Au, Je for
123 Jerrisbek, Ju for Juebek, Ki for Kielstau, Sa for Sankermark See, and Tr for the mainstream of
124 Treene. The numbers count along the longitudinal axis of rivers from the outlet to upstream. The
125 sampling points in close distance of lakes are not located in the lake, but rather are situated
126 systematically downstream following the lakes (Fig. 2).



127

128 **Fig. 2** The location of the Treene catchment and 59 sampling points in Schleswig-Holstein state of Germany.

129

130 *2.2. Land-cover pattern analysis (L)*

131 The catchment landscape is dominated by agricultural land-use. Around 50% of the area is
 132 covered by arable land, and around 30% by winter pasture. Soils in the north of the catchment
 133 (natural region of Angeln) are nutrient-rich and have a pH value ranging 6 to 7, while the pH of soils
 134 in the south-east of the catchment (Geest) varies from 4 to 5.5 (Tavares, 2006). Therefore, in the
 135 Angeln natural region, corresponding to Kielstau and Treene upstream catchments, agricultural fields
 136 are more common, meanwhile, in the southern Geest region (e.g., Jerrisbek and Juebek catchments)
 137 pastures constitute to a bigger part. The Treene catchment can be characterized by a slightly
 138 undulating landscape, with various depressions and lakes (Kiesel et al., 2010). Land-use data was
 139 provided by the Schleswig-Holstein State Bureau of Surveying and Geo-information
 140 (LVERMGEO-SH, 2012). The land-use analysis performed via ArcGIS software (Version 10.0, ESRI,

141 US) processing (Fig. A. 1). Eight land-use types were classified: agricultural land-generic (AGRL),
142 forests (FRST), rangeland (RNGE), industrial area (UIDU), urban areas (URMD), water bodies
143 (WATR), wetland (WETL) and winter pasture (WPAS). The upstream watershed area from each
144 sampling site was accumulated, and the land-use within this area was considered as the land-use
145 pattern affecting the sampling site. In this case, the sites located at downstream and near to the lakes
146 or cities presented a higher percentage of water bodies or urban area. For example, the sites located
147 on the upstream region of a city has none percentage of urban area land-use, while the downstream
148 sites have decreasing percentages along the river flow.

149 2.3. *Hydrological regime analysis (H)*

150 Except for water depth and velocity which were measured *in situ* at the sampling points
151 (velocity – using FlowSens Single Axis Electromagnetic Flow Meter, Hydrometrie, Germany), the
152 hydrological regime was mainly described by the indicators of hydrological alteration (IHA) (Olden
153 and Poff, 2003). The IHA, illustrating the hydrographic signatures from the duration, frequency,
154 timing magnitude and rate of flow events, are ecological relevant and can be calculated based on
155 daily discharge data and are also used in hydrology (Kiesel et al., 2017; Pool et al., 2017). The daily
156 discharge time series in this study based on the output of the ecohydrological SWAT model (Soil and
157 Water Assessing Tool). In this model application, the model discretization for the Treene catchment
158 resulted into 108 sub-basins. In a multi-site calibration, six hydrological stations which distributed in
159 the catchment were used to consider the spatial heterogeneity in reproducing discharge. The
160 modeling period subdivided into a calibration (2001 to 2005) and a validation period (2006 to 2016).
161 To evaluate the model performance, we used three well-known performance measures:
162 Nash-Sutcliffe Efficiency, Percent Bias and RSR (root mean square error divided by standard

163 deviation) (see Guse et al., 2015b for details). Daily modeled discharge values were used for
164 subbasins in which at least one sampling point located. At this moment, the sampling points were
165 related to the model results from the closest outlet of a sub-basin outlet. We used 57 hydrological
166 indices from IHA describing the traits of hydrological regime. The final hydrological matrix included:
167 magnitude, frequency, rate of flow events and *in situ* measurement (Table A.1).

168 2.4. Physicochemical factors analysis (P)

169 Physicochemical factors collection included a field *in situ* measurements and laboratory
170 measurement. Water temperature (WT), pH, electric conductivity (EC) and dissolved oxygen (DO)
171 of the surface water were measured *in situ* using Portable Meter (WTM Multi 340i and WTW Cond
172 330i, Germany) at each sampling site. Simultaneously, two water samples were stored in pre-cleaned
173 plastic bottles (500 ml each) for nutrient analysis in the laboratory. They were partially filtered
174 through GF/F glass microfiber filter (Whatmann 1825-047) for collecting the total suspended
175 substances. Both filtered and unfiltered samples were kept frozen at -20 °C until measurement
176 Concentrations of total phosphorus (TP), phosphate-phosphorus ($\text{PO}_4^{3-}\text{-P}$), ammonium-nitrogen
177 ($\text{NH}_4^+\text{-N}$), nitrate-nitrogen ($\text{NO}_3^-\text{-N}$), nitrite-nitrogen ($\text{NO}_2^-\text{-N}$), chloride (Cl^-) and sulfate (SO_4^{2-})
178 were measured using the standard methods of DEV (Deutsche Einheitsverfahren zur Wasser-,
179 Abwasser- und Schlammuntersuchung). Dissolved inorganic nitrogen (DIN) was defined as the
180 summation of nitrite-nitrogen ($\text{NO}_2^-\text{-N}$), nitrate-nitrogen ($\text{NO}_3^-\text{-N}$) and ammonium-nitrogen
181 ($\text{NH}_4^+\text{-N}$), and the nitrogen to phosphorus ratio (NPR) is the ratio between DIN and TP. In this study,
182 we chose DIN:TP ratio instead of the TN:TP ratio, since DIN:TP can better discriminate the N and P
183 limitation of phytoplankton (Bergström, 2010). Total suspended solids (TSS) were measured
184 according to Standard Operating Procedure for Total Suspended Solid Analysis (Federation and

185 Association, 2005).

186 2.5. *Phytoplankton collecting and processing*

187 Phytoplankton samples were quantitatively analyzed with a known concentration from the
188 subsurface (5 - 40 cm) water of the river by plankton net. Identification included two steps. First, soft
189 algae was identified in a Fuchs-Rosenthal chamber with an optical microscope (Nikon Eclipse
190 E200-LED, Germany) at $\times 400$ magnification, after employing the sedimentation method for the
191 samples (Sabater et al., 2008). Taxonomic identification was based on the references of Hu and Wei
192 (2006) and Burchardt (2014). Second, for further determination of the diatom species in the samples,
193 permanent slides were prepared after oxidization (using 5ml of 30% hydrogen peroxide, H_2O_2 , and
194 0.5ml of 1mol/l hydrochloric acid, HCl), and then 0.1 ml of the diatom-ethanol mix was transferred
195 on a 24 \times 24 mm coverslip. Diatoms were identified with the optical microscope (Nikon Eclipse
196 E200-LED, Germany) at $\times 1000$ under oil immersion, based on the key books by Bey (2013),
197 Hofmann et al. (2011) and Bak et al. (2012). Algal biomass was calculated using the approximation
198 of cell morphology to regular geometric shapes, assuming the fresh weight unit as expressed in mass,
199 where $1 \text{ mm}^3/\text{L} = 1 \text{ mg/L}$ (Huang, 2000; Wetzel and Likens, 2013). We then calculated the relative
200 biomass of every species as their abundance. Based on the criteria proposed by Reynold (2002),
201 species that contributed more than 5% of the total biomass were sorted into functional groups, and
202 the phytoplankton functional classification was done according to Reynolds et al. (2002), Borics et al.
203 (2007) and Padisák et al. (2009). Species not mentioned in the references were assigned to a group
204 according to their morphological and ecological characteristics and the environmental conditions
205 prevailing during their greatest occurrence (Devercelli, 2006). In this study, we specially focused on
206 the functional group **M** (species mainly from genus: *Microcystis* consisted by *Microcystis*

207 *aeruginosa*, *M. wesenbergii*, *M. viridis*) and **TB** (mainly composed of benthic Pennales, typically
208 genus such as: *Navicula*, *Nitzschia*, *Gomphonama*, *Fragilaria*). From the references, this two group of
209 algae have different strategies, preference of living, and they present two characteristic conditions:
210 highly lotic river sections with **TB** dominance, while **M** dominance in eutrophicated lentic habitats
211 (Borics et al., 2007).

212 2.6. Statistical analysis

213 For achieving the best performance, the phytoplankton functional groups abundance matrix was
214 Hellinger transformed (Legendre and Gallagher, 2001; Legendre and Legendre, 2012). Firstly, the
215 analysis of PERMANOVA was conducted for testing the difference of PFG composition (function:
216 *adonis* of the R-package: *vegan*). In the meanwhile, the three sets of abiotic variables (i.e., L, H and
217 P) collinearity was tested when using all variables in the model of explaining variations of
218 phytoplankton communities (function: *cor* of the R-package: *stats*). Afterwards, a forward selection
219 (Blanchet et al., 2008) was carried out to choose a parsimonious subset of explanatory variables
220 (function: *forward.sel* of the R-package: *adespatial*), and then the multivariate community structure
221 was modeled under variation partitioning analysis (Borcard et al., 1992) (function: *rda*, *varpart* of
222 the R-package: *vegan*). We then constructed structural equation model (SEM) (Westland, 2016) for
223 quantitative evaluation of the relationship between human and natural stressors on PFG composition,
224 as well as the potential effect of land-use and hydrology on in-stream nutrients. We used
225 reduced-multidimensional data rather than original PFG matrices in the analysis since SEM can only
226 handle one-dimensional variables. Nonmetric multidimensional scaling (NMDS) ordination was
227 applied to produce reduced-dimensional data of PFG composition, measured as the first NMDS axis
228 scores for each site (i.e., NMDS1) to represent the main condition of the whole phytoplankton

229 community composition (function: *metaMDS* of the R-package: *vegan*). To achieve the SEM, we
230 first proposed the conceptual model for the three bio-indicators (PFG: **TB**, **M** and NMDS1). The
231 conceptual model included all possible pathways between the response variables and their key
232 abiotic variables, which identified by general linear model (function: *glm* and *step* of the R-package:
233 *stats*). In addition to the direct pathways, we also checked the indirect ones to see if variables exerted
234 further effects via mediation variables. From the initial model, we then specified the pathways. All
235 non-significant paths were eliminated stepwise until all remaining paths were significant related to
236 the response variables. Standardized path coefficients were calculated for each pathway for a better
237 comparable in the SEM. The overall final model fit was evaluated with root mean square error of
238 approximation (RMSEA), and the comparative fit index (CFI). RMSEA approach to 0, and CFI close
239 to 1.0 indicate a good fit of the model (Grace, 2006). In this study, SEM was constructed by *sem* in
240 the R-package: *lavaan*.

241 All analyses were conducted with the R software (version 3.5.1, R Development Core Team,
242 2018).

243 **3. Results**

244 *3.1. Description of watershed hydrological regime and physicochemical condition*

245 From the model outputs, the indicators of hydrological alteration varied across the two studied
246 periods. Water flow has a higher magnitude and fluctuation in the high flow (December) than in the
247 low flow period (September) (e.g., at the outlet of the catchment Tr01, Fig. A. 2). Some other typical
248 hydrological indices, for instance, H20 which represents the skewness of seven days of discharge,
249 has an opposite spatial trend during these two periods (Fig. A. 3). The index H20 ranges from -0.775

250 to 2.439. Most values of H2O were positive with a range of 0.043 to 2.439. They were negative
251 skewed only in the upstream of the main river (specifically sites: Tr16 - Tr22) in high flow period,
252 and tributary Jerrisbek during dry season. We observed a relatively high value in the tributary
253 Jerrisbek during the wet season (with an average value of 2.038). The key hydrological indicators
254 were summarized after pre-selection excluding the ones with significant multi-collinearity (Table 1).

255 The local physicochemical variables measured *in situ* varied considerably among different
256 hydrological periods and sub-catchments (Fig. A. 4). Firstly, the difference of water temperature
257 (WT) in two seasons was remarkable, and mean values in December 2014 and September 2015 were
258 5.69 °C and 1424 °C, respectively (Table 1). The value of pH followed a similar pattern in different
259 sub-basins, while mean value in September was slightly higher than in December ($8.084 > 7.487$,
260 Table 1). The concentrations of ammonium-nitrogen ($\text{NH}_4^+\text{-N}$) showed a higher concentration in
261 December compared to September (average: $0.307 \text{ mg/L} > 0.158 \text{ mg/L}$, Table 1, also see more
262 details in Fig. A. 4). Conversely, nitrate-nitrogen ($\text{NO}_3^-\text{-N}$) concentration showed lower in December
263 than in September (average: $3.551 \text{ mg/L} < 9.243 \text{ mg/L}$, Table 1, also see more details in Fig. A. 4).
264 Dissolved inorganic nitrogen (DIN) followed a similar pattern as nitrate-nitrogen. We observed a
265 decreasing trend of phosphate-phosphorus ($\text{PO}_4^{3-}\text{-P}$) concentration while an increasing trend of
266 nitrogen to phosphorous ratio (NPR) along the mainstream of Treene (Tr) in September 2015. Except
267 for the sub-basin Juebek, the average value and fluctuation of NPR in September was higher than in
268 December (Fig. A. 4). The concentrations of $\text{NH}_4^+\text{-N}$ and $\text{PO}_4^{3-}\text{-P}$ in sub-basin Kielstau were always
269 significantly higher than other sub-basins at both investigation periods. Generally, the nutrient
270 contents such as $\text{NO}_3^-\text{-N}$, DIN and NPR in wet season were lower than in dry season (Table 1).

271 Table 1 Summary of important hydrological (H), and physicochemical (P) variables with their codes and
 272 descriptions in this study. Variables with significant multicollinearity (spearman correlation coefficient greater or
 273 equal to 0.75) are excluded from the table.
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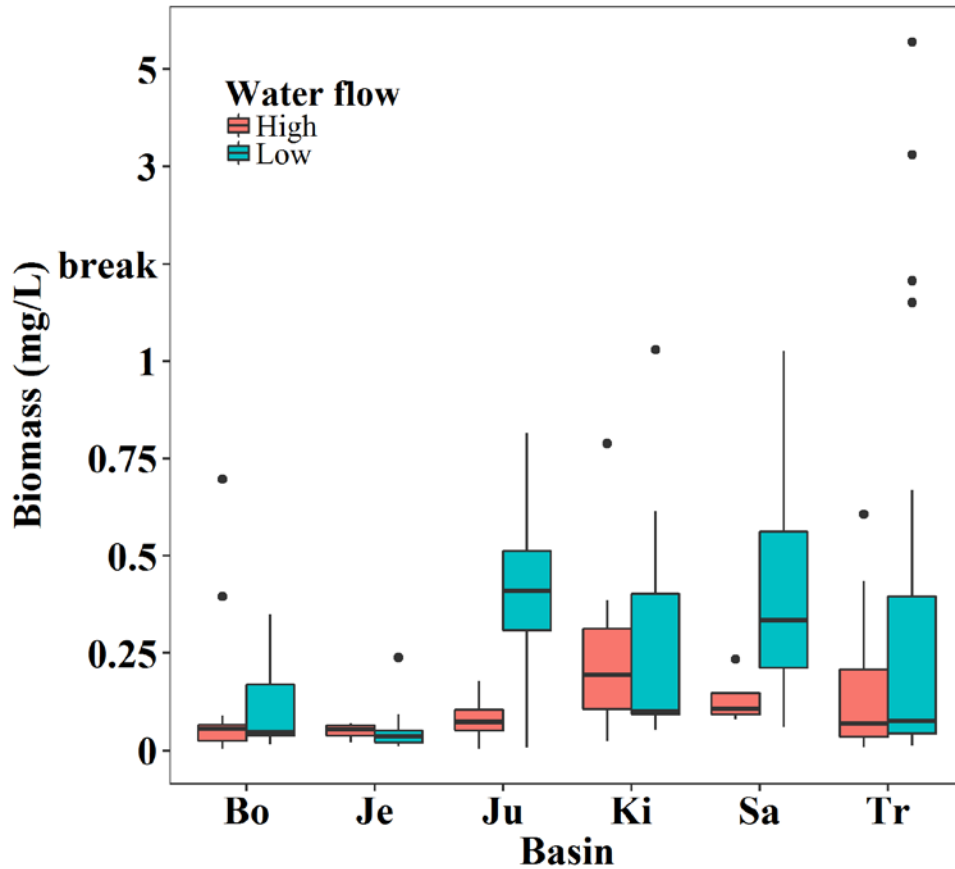
Variables						
Code	Unit	Description	High flow		Low flow	
			Mean	SD	Mean	SD
H		Hydrological parameter				
H01	m ³ /s	Discharge at the sample day	2.274	4.38	0.299	0.52
H20	-	Skewness of 7 days' discharge	0.955	0.686	0.814	0.696
H36	-	Skewness of 30 days' discharge	1.143	0.636	1.428	0.435
H41	days	Low flood pulse count 30 days	13.373	10.257	22.373	3.737
H42	days	High flood pulse count 3 days	1.492	1.467	0	0
H55	-	Rate of change in 7 days	-0.038	0.089	-0.017	0.032
P		Physicochemical parameter				
WT	°C	Water temperature	5.69	1.581	14.237	1.619
pH	-	pH	7.487	0.474	8.084	0.407
NH4	mg/L	Ammonium-nitrogen (NH ₄ ⁺ -N)	0.307	0.276	0.158	0.266
NO3	mg/L	Nitrate-nitrogen (NO ₃ ⁻ -N)	3.551	1.772	9.243	6.784

NO2	mg/L	Nitrite-nitrogen (NO ₂ ⁻ -N)	0.02	0.019	0.054	0.119
TP	mg/L	Total phosphorus	0.225	0.116	0.211	0.209
PO4	mg/L	Phosphate-phosphorus (PO ₄ ³⁻ -P)	0.077	0.058	0.072	0.109
NPR	-	Nitrogen to phosphorus ratio	21.002	14.926	79.739	67.715
SO4	mg/L	Sulfate (SO ₄ ²⁻)	31.819	10.923	44.087	14.572

275

276 3.2. Variation of phytoplankton assemblages

277 We observed 396 algal taxa from the 118 samples and they were classified into 21
278 phytoplankton functional groups (PFGs). Among them, there were 16 groups in December 2014,
279 while 19 groups in September 2015. The PERMANOVA analysis showed a significant dissimilarity
280 of phytoplankton functional groups composition both temporal (high water flow period and low
281 water flow period) and spatial (sub-basins). We also found that biomass increased significantly from
282 high flow to low flow period in the sub-basins of Tr, Sa and Ju, while fewer changes occurred in the
283 sub-basins of Bo, Je and Ki (Fig. 3). In both hydrological periods, functional group **TB** constituted a
284 high portion in sub-basins: Bo (87% - Dec. 2014, 68% - Sep. 2015) and Je (98% - Dec.2014, 64% -
285 Sep. 2015). In contrast, the functional group **M** percentage increased in the sub-basin of Tr, Ki, Sa
286 and Ju from December to September (Table A. 2).



287

288 **Fig. 3** Average biomass in different basins (Bo represents for sub-basin Bollingstedter Au, Je for Jerrisbek, Ju for
 289 Juebek, Ki for Kielstau, Sa for Sankermark See and Tr for mainstream of Treene)

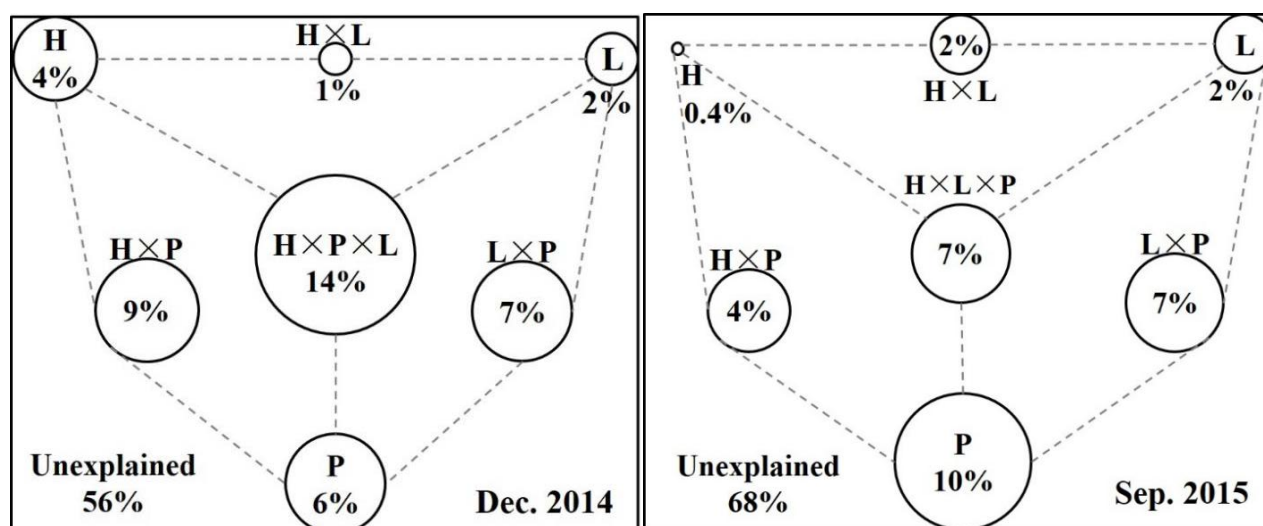
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291 *3.3. Effect of abiotic factors on phytoplankton functional groups*

292 *3.3.1. Relationship described by variation partitioning*

293 In wet season (Dec. 2014), there were 3 L, 5 H and 5 P variables selected by a forward selection
 294 (supporting information Table A. 3). All the groups showed significant relationships with PFGs (by
 295 *anova* function in R, $p < 0.001$). According to variation partitioning analysis, the three sets could
 296 explain 44% of the variation in PFGs (Fig. 4). The variations purely explained by H, L and P were
 297 4%, 2% and 6%, respectively, while the shared fraction of the three variables was 14%. In general,
 298 the joint contribution by H and P (H×P, 9%) was higher than those by H×L (2%) and P×L (7%). In
 299 the dry season (Sep. 2015), there were 2 L, 1 H and 6 P variables selected by a forward selection

300 (Table A. 3). The PFGs variation was mostly related to the effect of P (their pure effect explained
 301 10% of the variation), whereas the pure effect of H was the least (only accounted for 0.4%). The
 302 variation partitioning also showed that both wet and dry periods had similar explanation fraction of
 303 land cover (Dec. 2014: 2%; Sep. 2015: 2%).



304
 305 **Fig. 4** Contributions of the hydrological (H), land-cover (L) and physicochemical (P) variables to the variances in
 306 phytoplankton functional groups (PFGs). Each diagram represents a given biological variation partitioned into the
 307 pure effects of H, L and P (i.e. when removing the variations caused by other two factors), interaction between any
 308 two variables (HxL, LxP, HxP), interaction of all three factors (HxLxP) and unexplained variation (total variation
 309 = 100). The analysis includes two scenarios: left: illustrate the situation in December of 2014 (n=59), right: show
 310 the results in September of 2015 (n=59).

311

312 3.3.2. Relationships described by structural equation model

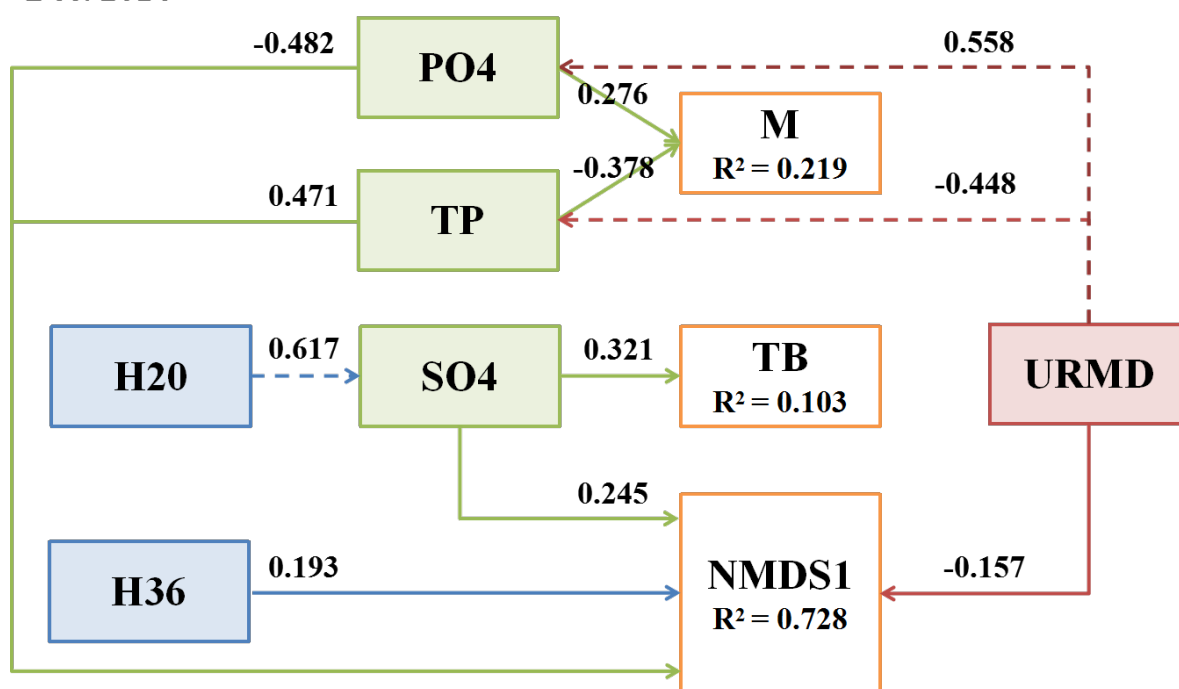
313 From the general linear model results (supporting information Table A. 4), we got a general
 314 idea about the effects of the multiple stressors. For further investigation of the causal relationship
 315 between the abiotic and biotic variables, we fitted structural equation models (SEMs) to infer the
 316 direct and indirect effects of specific abiotic variables on bio-indicators (indicated by NMDS1 and
 317 phytoplankton functional groups **M**, **TB**) (Fig. 5).

318 In high flow period (Dec. 2014), SEMs indicated that group **M** was directly governed by

319 PO₄³⁻-P and TP ($\beta=0.276, 0.378$, respectively, standardized coefficient), while indirectly affected by
 320 URMD via PO₄³⁻-P and TP ($\beta=0.558, -0.448$, respectively, standardized coefficient). The
 321 hydrological variable H2O (skewness of 7 days' discharge) was mediated through sulfate (SO₄²⁻)
 322 effect on **TB**. The SEM explained 72.8% of the variation of NMDS1. The explanations come from
 323 H36 (skewness of 30 days' discharge), PO₄³⁻-P, TP, SO₄²⁻ directly, and URMD indirectly via H2O.

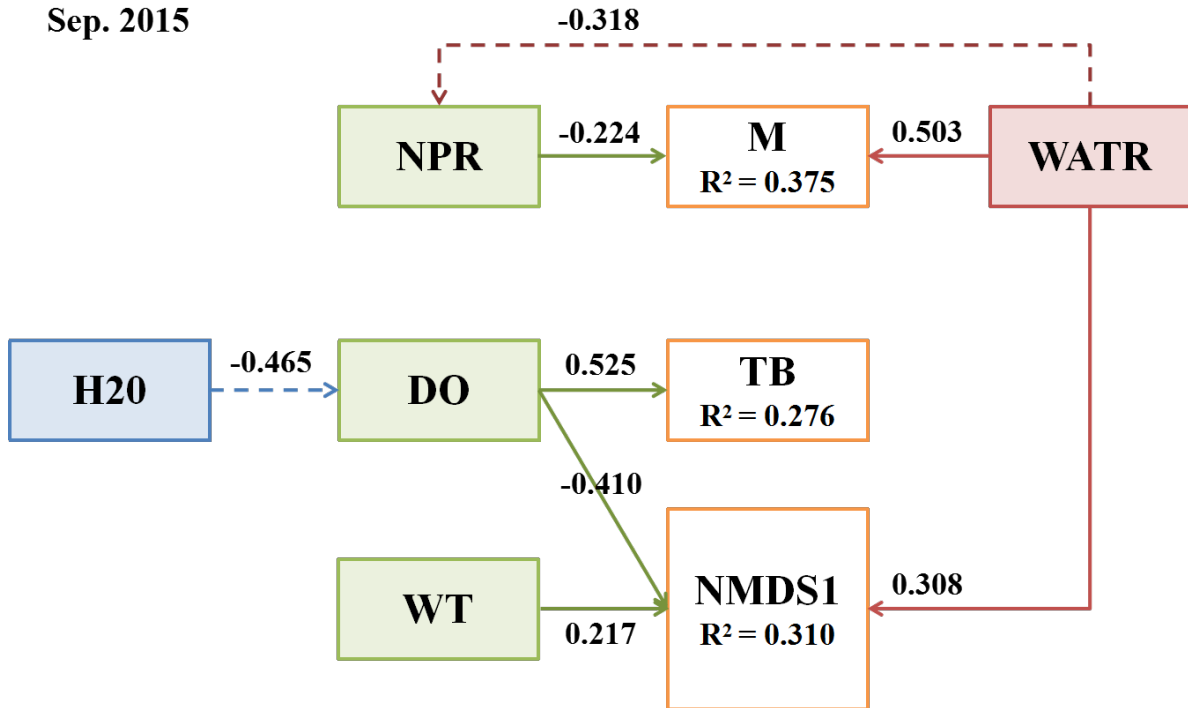
324 During the low flow period (Sep. 2015), the strongest relationship ($\beta=0.525$, standardized
 325 coefficient) was observed in the SEM analysis between functional group **TB** and dissolved oxygen
 326 (DO). H2O exerted an indirect effect on **TB** via DO ($\beta=-0.465$, standardized coefficient). Area
 327 covered by water (WATR) appeared as the key variable for the group **M**, directly ($\beta=0.503$,
 328 standardized coefficient) and indirectly through NPR ($\beta=-0.318$, standardized coefficient). The SEM
 329 explained 31% of the community variation (represented by NMDS1) was negatively affected by DO,
 330 and positively affected by WT and WATR.

Dec. 2014



331

Sep. 2015



332

333 **Fig. 5** Structural equation models embody the causal relationships between hydrological regime (H), land-use
 334 pattern (L), physicochemical condition (P) and phytoplankton bio-indicators. Solid arrows represent direct paths
 335 ($p < 0.05$), and dashed arrows represent indirect paths. The values corresponding to the path coefficients have
 336 standardized effect size. The models are evaluated using R^2 . The hydrological parameters are in blue squares,
 337 physicochemical parameters in green squares, land-use parameters in red squares, and the orange squares represent
 338 phytoplankton functional groups (M, TB) and community index (NMDS1). The analysis includes two scenarios:
 339 the upper one illustrates the situation in December of 2014 ($n=59$), the lower one shows the results on September of
 340 2015 ($n=59$). The environmental variables included: H20: skewness of 7 days' discharge (including the sampling
 341 day); H36: skewness of 30 days' discharge (including the sampling day); WATR: area covered by water; URMD:
 342 urban area with residential-medium density; PO4: phosphate- phosphorus; TP: total phosphorus; SO4: sulfate; NPR:
 343 nitrogen to phosphorus ratio; DO: dissolved oxygen; WT: water temperature.

344

345 4. Discussion

346 4.1. Response of TB to multiple stressors

347 We observed a higher contribution of hydrological regime during high water period (Fig. 4).

348 The result demonstrated the importance of flow regime in shaping phytoplankton community

349 population and composition, which was also supported by previous results in the study region (Qu et

350 al., 2018a). Among the hydrological variables, skewness of discharge in 7 days (H20) emerged as the
351 key factor in shaping the pattern of **TB** both in wet and dry periods of the study (Fig. 5). The index
352 H20 provided two opposite trends across different hydrological periods. One river could present two
353 types of hydrograph on skewness depending on the hydrological conditions, such as in the river
354 Piquiri, South Brazil, where small (large) floods present positive (negative) skewness (Fleischmann
355 et al., 2016). The maximum value of skewness of flow in the studied catchment was observed in the
356 tributary Jerrisbek during the wet season. As we know, the Jerrisbek tributary is in the flatter hill
357 region. Additionally, the region had a higher share of forest and pasture, which turned to interfluvial
358 wetland during flooding, which leads to low flood wave attenuation (Junk et al., 2011). A larger
359 skewness implied higher discharge and runoff variability (Poff and Zimmerman, 2010).

360 Higher percentage of **TB** was observed in the tributary of Jerrisbek compared to other areas of
361 the river (Table A. 2). The PFG **TB** as typical potamal phytoplankton better dominated in high flow
362 condition of small rivers (Stanković et al., 2012). The result support the hypothesis that increased
363 water discharge can trigger higher share of **TB** in the community, due to their relatively high
364 tolerance to flushing and turbulence (Borics et al., 2007). Phytoplankton communities were
365 co-dominated by planktic and benthic algae. The proportion of silicified benthic diatoms increased
366 when water residence time decreased (Beaver et al., 2013). Consistent with this, Wang et al. (2018)
367 demonstrated that the community composition of benthic diatoms in river plankton was not
368 randomly distributed. Additionally, it could be related to algal hydrological constraints based on their
369 morphological and functional traits aspects in a meaningful way. Likewise, B-Béres et al. (2016) also
370 emphasized that the trait of guild was a sensitive indicator for the environmental conditions in the

371 lowland rivers and streams.

372 Furthermore, we also observed a significant positive relationship between sulfate and benthic
373 diatoms. The results revealed the possible importance of sulfate for diatoms growth. Based on a
374 controlling experiment, sulfate is likely to be a promotion factor, which would be a benefit to the
375 photosynthetic characteristics of benthic diatoms (Lengyel et al., 2015).

376 *4.2. Response of **M** to multiple stressors*

377 During high flow period, domestic area (URMD) is selected as an indirectly structuring factor
378 to **M** via essential nutrient phosphorus (Droop, 1974), and the soluble phosphate was positive related
379 with **M** (Fig. 4). It was surprisingly against with our hypotheses that the key factor would be the
380 major land-use of agriculture. However, the results are consistent with other previous findings. In the
381 Taizi River basin (China), they reported the pollution sources from built-up land areas were existing
382 in both rainy and dry seasons as a point source, where the population is dense and industrial activities
383 are intensive (Bu et al., 2014). It was reported that the urbanized land-use had better production of
384 water quality than agricultural land-use, indicating that urbanized land-use is the primary contributor
385 to degraded water quality rather than agricultural land-use (Baker, 2003; Schoonover and Lockaby,
386 2006). Additionally, it was reported a continuous high supplement of nitrogen, while controlled
387 phosphorus in the rural area (Mischke et al., 2011). The condition of phosphorous showed as an
388 important nutrient factor, especially for the growth of **M**.

389 During the low flow period, the area covered by water body (WATR) appeared as another key
390 factor for controlling the functional group **M** with a direct positive relationship (Fig. 5). The result
391 indicated that the lake act as a source of *Microcystis* rather than a sink of purification in the

392 catchment. The biomass of the percentage of **M** was the highest on the sites after lakes. On the
393 contrary, Lake Durowskie (west Poland), which is also located in a farmland-dominant catchment,
394 has suffered inflow with severe cyanobacteria blooms by the linked Struga Gołaniecka River.
395 However, the outflow water quality recovered by three restoration methods in the lake (Gołdyn et al.,
396 2014; Kowalczywska-Madura et al., 2018). As a typical planktic group, **M** has been regarded as a
397 group with the preference of small, eutrophic lacustrine habitat (Borics et al., 2007), while
398 sensitivities to flushing and low light (Reynolds et al., 2002). In lentic area, high phytoplankton
399 bio-volume may dominate by bloom-forming cyanobacteria occurring due to high water residential
400 time (Rangel et al., 2016). The upstream dam resulted in lake type eutrophic, epilimnetic *Microcystis*
401 dominance, while replaced by benthic diatoms in the middle sections of the River Loire (France) in
402 late summer (Abonyi et al., 2012).

403 Moreover, we also observed that the ratio of nitrogen to phosphorous was directly negative
404 related to group **M** during the dry season (Fig. 5). On one hand, the result showed that the content of
405 phosphorous acted as the shaping factor for *Microcystis* development (Schindler et al., 2016). On the
406 other hand, the negative relationship between NPR and **M** consistent with the previous studies that
407 cyanobacteria bloom was favored by low NPR (Orihel et al., 2015; Smith, 1983).

408 Finally, we are specially facing a high biomass of **M** on the low flow period, when local farmers
409 fertilized crops and used pesticides more extensively in the study region (Ulrich et al., 2018).
410 Cyanobacteria had been noticed by higher tolerance to herbicides than other phytoplankton taxa
411 (Bérard et al., 1999), particularly under status of enhanced nutrient supply (Harris and Smith, 2016),
412 indicating pesticides might potentially stimulated the dominance of **M** during low flow period.

413 *4.3. Response of phytoplankton community to multiple stressors*

414 During the high flow period, the phytoplankton community pattern had a higher influence from
415 hydrological indexes compare to the dry period (Fig. 5). The hydrological regime mainly contributed
416 to the low biomass and high similarity of the phytoplankton composition during the high flow period,
417 due to the high connectivity and extensive dispersal stochasticity (Rodrigues et al., 2018). On the
418 contrary, there was a relative high level of total biomass during the dry season (Fig. 3), and a high
419 share of contribution from physicochemical condition in structuring the phytoplankton community
420 (Fig. 4). Firstly, it can be expected a light limitation in winter months in the Northern Hemisphere. In
421 addition, we could infer even higher light availability in the dry season due to the low water level
422 (Nõges et al., 2016). Moreover, water temperature had a positive contribution to the phytoplankton
423 population development in September (Fig. 5). This phase of the year had more suitable temperature
424 for algae growth compared to December. Warm autumn water temperatures combined with
425 anthropogenic eutrophication attributed to the cause of riverine algal blooms during drought period
426 (Bowling et al., 2016).

427 Overall, our findings suggest that the impacts of flow regime, land-use pattern, physicochemical
428 condition and their potential interactions on riverine PFGs (**TB**, **M** and the community) varied
429 greatly across hydrological periods. However, traditional biomonitoring campaigns and management
430 practices, which focused on improving local abiotic variables to increase local biodiversity, were
431 often based on one-time sampling data and fairly ignored the potential bias resulting from temporal
432 variations, particularly different hydrological periods (Stubbington et al., 2017b). The designation
433 was probably one of the reasons that have resulted into many problems and delays in implementation
434 of recent international water framework directive policies such as EU WFD (Voulvoulis et al., 2017).

435 We therefore advocate that sampling programs and analyses that target future environmental policies
436 should take different hydrological periods into account (Stubbington et al., 2017a). Furthermore,
437 studies address that detailed causal relationships between changing environment and aquatic
438 organisms could significantly improve our basic understanding of ecological responses to multiple
439 stressors. Sustainable watershed management requires holistic approaches that assess collective
440 biotic and abiotic data to better predict the impacts of anthropogenic activities and climate change on
441 aquatic ecosystems (Dudgeon et al., 2006).

442 **5. Conclusions**

443 In this study, the phytoplankton biomass and functional groups composition vary
444 spatiotemporally during two contrasting seasonal hydrological periods. Their responses to
445 physicochemical conditions, hydrological regime and land-use pattern are different during different
446 hydrological periods. we detect that:

447 (1) The hydrological regime contributes more during the high flow period in structuring the
448 phytoplankton community. The skewness of 7 days discharge emerged as a key driver of
449 hydrological regime, and it always had an indirect effect on functional group **TB** during two
450 hydrological periods.

451 (2) Anthropogenic stressor by urban land-use acted as a critical driver for functional group **M**
452 especially during high flow period. The group **M** was directly related to phosphorous
453 relevant indicators indicating its sensitivity to the concentration of phosphorous. The area
454 with a higher share of water bodies was also a key cause factor during the low flow period,
455 guide the lacustrine zone as its origin.

456 Our results provide evidence that, for a more comprehensive and accurate understanding of the
457 aquatic status, we should also take consideration of different hydrological periods. Secondly, we
458 recommend functional groups as effective indicators of phytoplankton dynamics to simplify the
459 pattern and seize the key issue to achieve a better knowledge of the relationship between
460 phytoplankton and multiple environmental stressors. Climate change and anthropogenic activities,
461 for instance altering flow regime and land-use pattern, act as an important ecological filter of riverine
462 phytoplankton community via physicochemical conditions. Further studies across lake-river
463 continuums are needed to enhance our understanding toward incorporating hydrological regime,
464 land-use pattern, as well as physicochemical conditions into investigation designs, which would
465 enable us to keep the pace with increasing demand for sustainable watershed management. Structure
466 equation model disentangled the contribution of multiple stressors. It would be an outperforming
467 method to generalize results, identify common patterns, and predict response of phytoplankton to
468 environmental changes in future studies.

469 **Acknowledgments**

470 This study has funded by German Research Foundation (Deutsche Forschungsgemeinschaft DFG)
471 grants (FO 301/15-1, FO 301/15-2, WU 749/1-1, WU 749/1-2, and the project GU 1466/1-1
472 Hydrological Consistency in Modelling). There is financial support scholarship by AIAS CO-FUND
473 funding (Naicheng Wu) and China Scholarship Council (CSC) (Yueming Qu). We thank: Dr. Fuqiang
474 Li, Zhao Pan and other friends for their support during the field campaigns, and the laboratory team
475 of the Department of Hydrology and Water Resources Management of the Christian Albrechts
476 University of Kiel for carrying out the water quality analysis. We also thank Ms. Rebecca Pederson for

477 help to revise the language problems. The valuable comments of two anonymous reviewers have
478 improved the manuscript greatly.

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